

Fusion of Inactivated Cas9 to *FokI* Nuclease Improves Genome Modification Specificity

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SUPPLEMENTARY RESULTS

Development of the XTEN-based linker

The XTEN protein was originally designed to extend the serum half-life of translationally fused biologic drugs by increasing their hydrodynamic radius, acting as protein-based functional analog to chemical PEGylation.¹ Since XTEN is chemically stable, non-cationic, non-hydrophobic, and predicted to adopt an extended, unstructured conformation, we hypothesized that an XTEN-based linker could functional as a stable, inert linker sequence for fusion proteins. The sequence of the XTEN protein tag from E-XTEN was analyzed, and repeating motifs within the amino acid sequence were aligned. The sequence used in the *FokI-dCas9* fusion construct *FokI-L8* (**Supplementary Figure 4a**) was derived from the consensus sequence of a common E-XTEN motif, and a 16-residue sequence was chosen from within this motif to test as a *FokI-dCas9* linker.

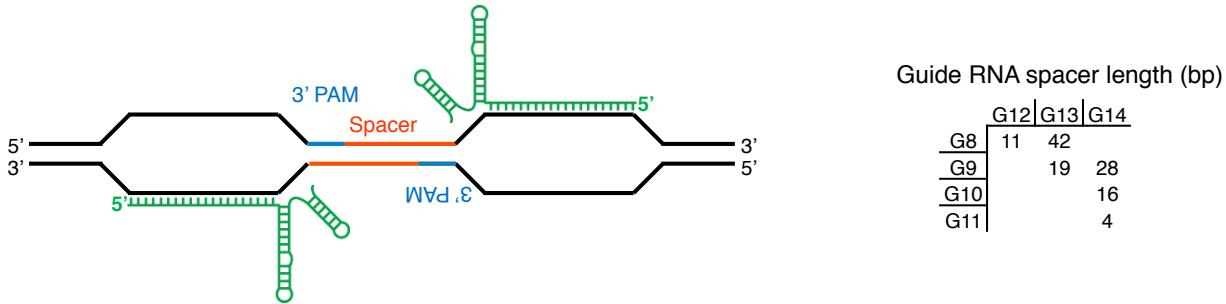
Additional NLS-*FokI* linker variants

In addition to assaying linkers between the *FokI* domain and dCas9 in the NLS-*FokI-dCas9* architecture, we also tested four linker variants between the N-terminal NLS and the *FokI* domain (**Supplementary Figure 4a**). Although a NLS-GSAGSAAGSGEF-*FokI-dCas9* linker exhibited nearly 2-fold better GFP gene modification than the other NLS-*FokI* linkers tested when a simple GGS linker was used between the *FokI* and dCas9 domains (**Supplementary Figure 4b**), the GSAGSAAGSGEF linker did not perform substantially better when combined with the XTEN linker between the *FokI* and dCas9 domains.

Sensitivity limit of off-target cleavage assays

The sensitivity of the high-throughput sequencing method for detecting genomic off-target cleavage is limited by the amount genomic DNA (gDNA) input into the PCR amplification of each genomic target site. A 1 ng sample of human gDNA represents only ~330 unique genomes, and thus only ~330 unique copies of each genomic site are present. PCR amplification for each genomic target was performed on a total of 150 ng, 300 ng, or 600 ng of input gDNA, which provides amplicons derived from at most 50,000, 100,000 or 200,000 unique gDNA copies, respectively. Therefore, the high-throughput sequencing assay cannot detect rare genome modification events that occur at a frequency of less than 1 in 50,000 (0.002%), less than 1 in 100,000 (0.001%), or less than 1 in 200,000 (0.0005%), respectively.

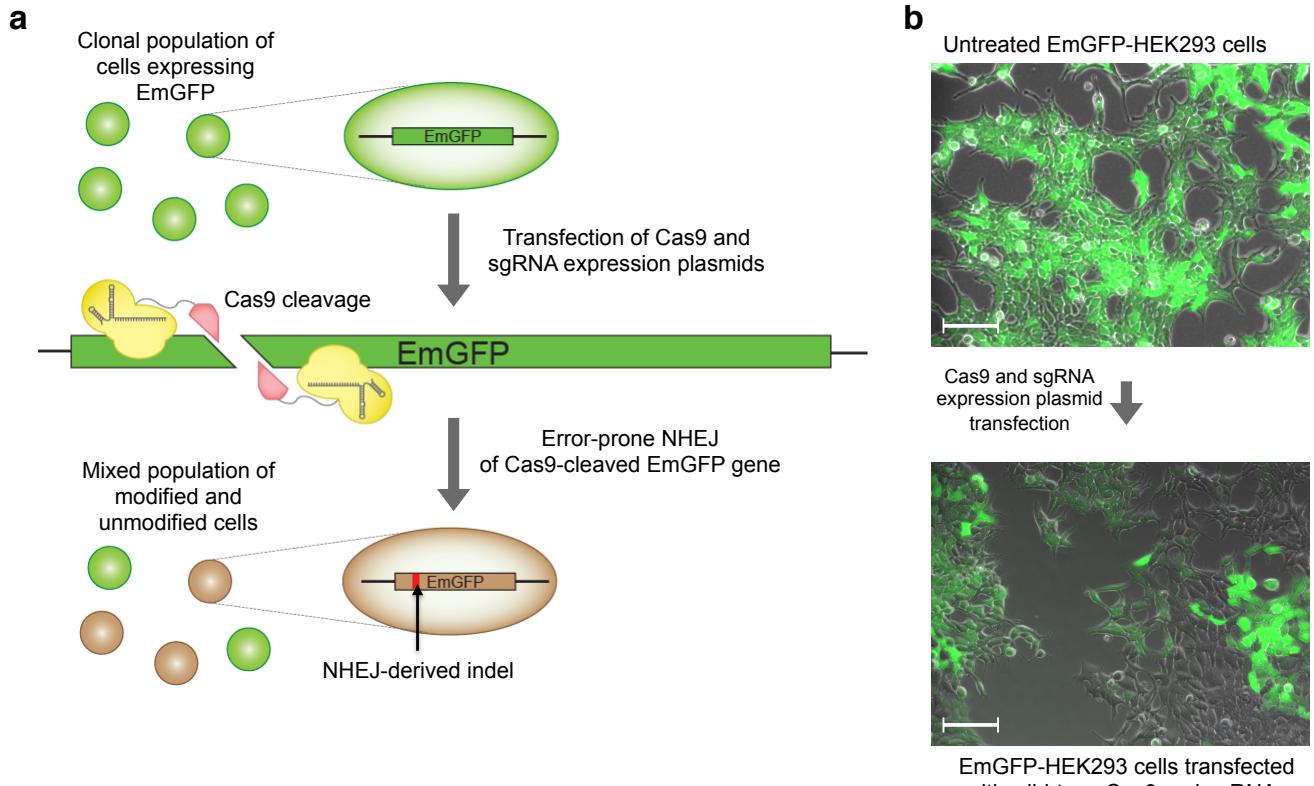
Orientation B:



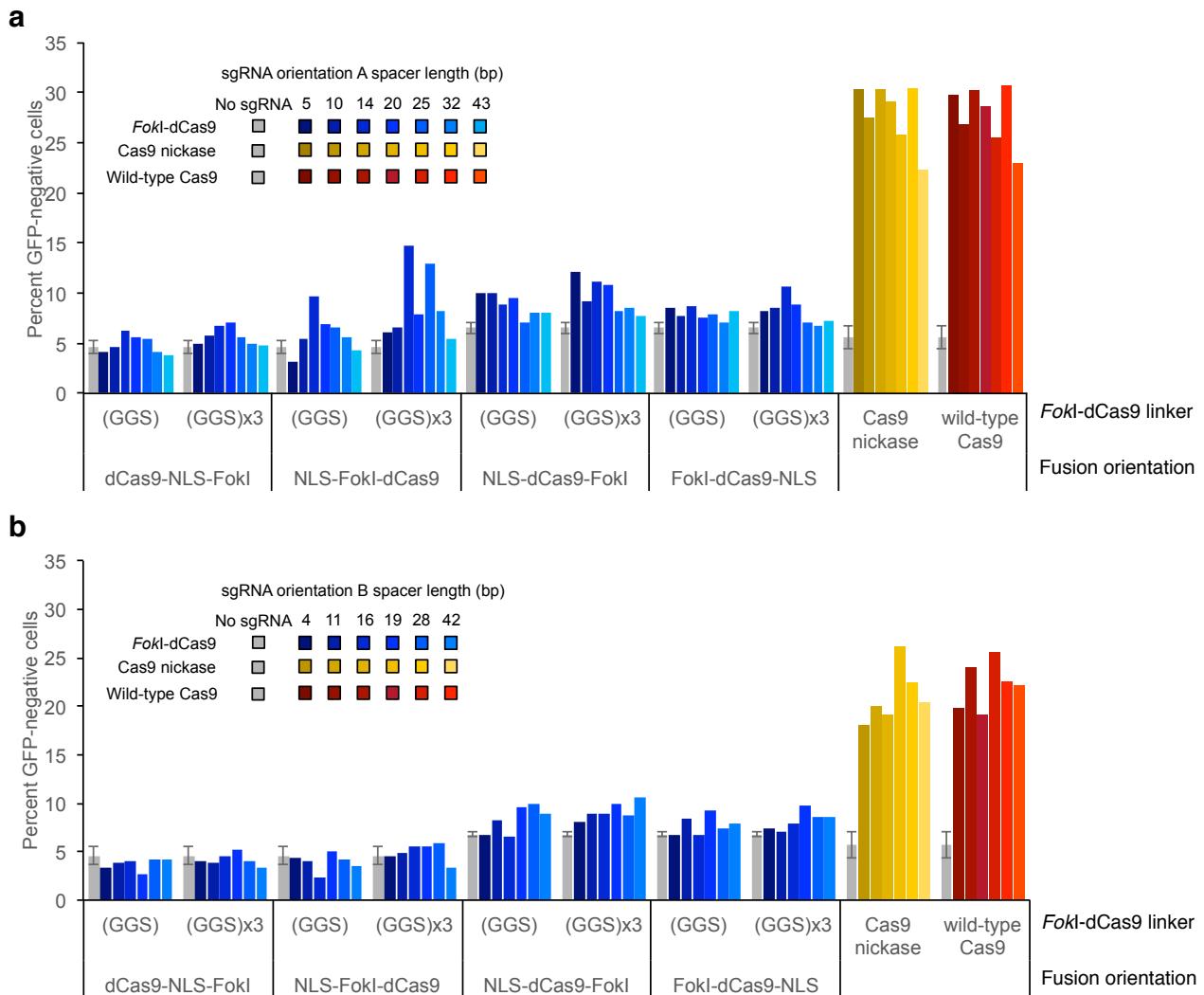
Guide RNA sequences within EmGFP (bp 297-388)

5' -GGAGCGCACCATTTCTCAAGGACGACGGCAACTACAAGACCCGGCCGAGGTGAAGTTCGAGGGGACACCCCTGGTGAACCGCATCGAGCTGAAGGGCATCG-3'
 G8 ACCATTTCTCAAGGACGAGG
 G9 CAACTACAAGACCCGGCCGAGG
 G10 CCGCGCCGAGGTGAAGTTCGAGG
 G11 GAAGTTCGAGGGCAGACACCCCTGG
 G12 CCCGCGCCGAGGTGAAGTTCGAG
 G13 CCTGGTGAACCGCATCGAGCTGA
 G14 CCGCATCGAGCTGAAGGGCATCG

Supplementary Figure 1. Target DNA sequences in a genomic GFP gene. Seven sgRNA target sites were chosen to test *FokI-dCas9* candidate activity in an orientation in which the PAM is adjacent from the cleaved spacer sequence (orientation B). Together, these seven sgRNAs enabled testing of *FokI-dCas9* fusion variants across six spacer lengths ranging from 4 to 42 bp.



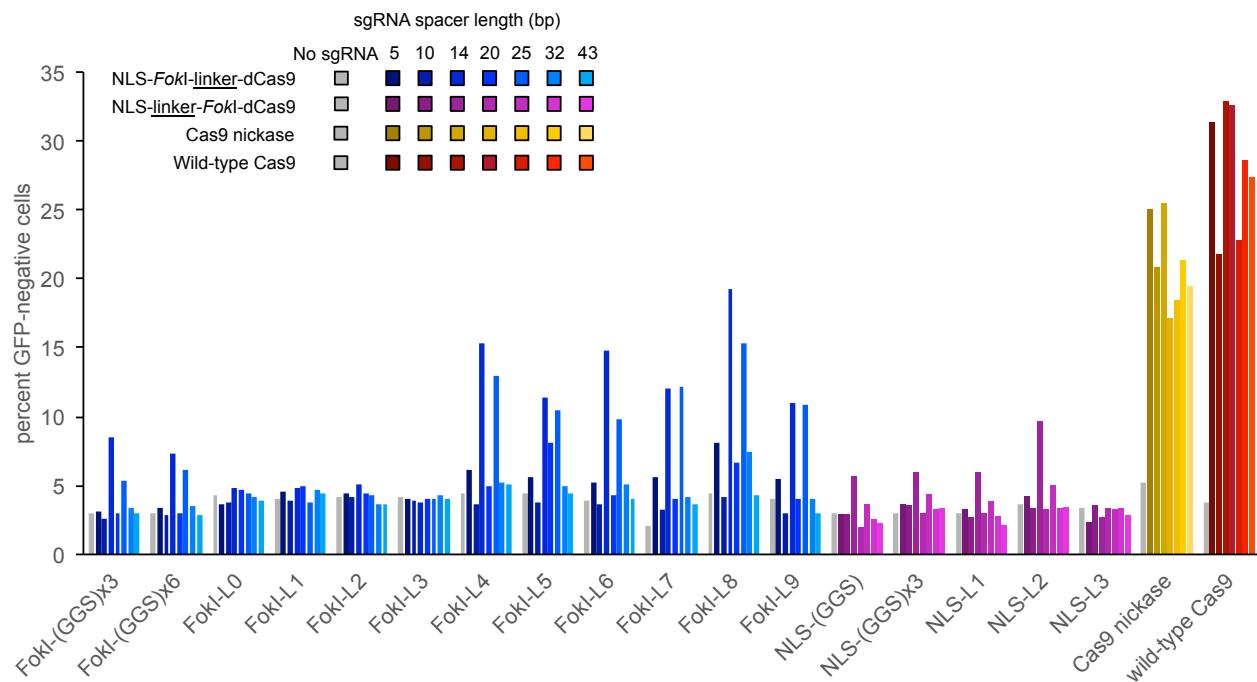
Supplementary Figure 2. GFP disruption assay for measuring genomic DNA-modification activity. (a) A HEK293-derived cell line constitutively expressing a genomically integrated EmGFP gene was used to test the activity of candidate *FokI*-dCas9 fusion constructs. Co-transfection of these cells with appropriate nuclease and sgRNA expression plasmids leads to dsDNA cleavage within the EmGFP coding sequence, stimulating error-prone NHEJ and generating indels that can disrupt the expression of GFP, leading to loss of cellular fluorescence. The fraction of cells displaying a loss of GFP fluorescence is then quantitated by flow cytometry. (b) Typical epifluorescence microscopy images at 200x magnification of EmGFP-HEK293 cells before and after co-transfection with wild-type Cas9 and sgRNA expression plasmids. White scale bars represent 200 μ m.



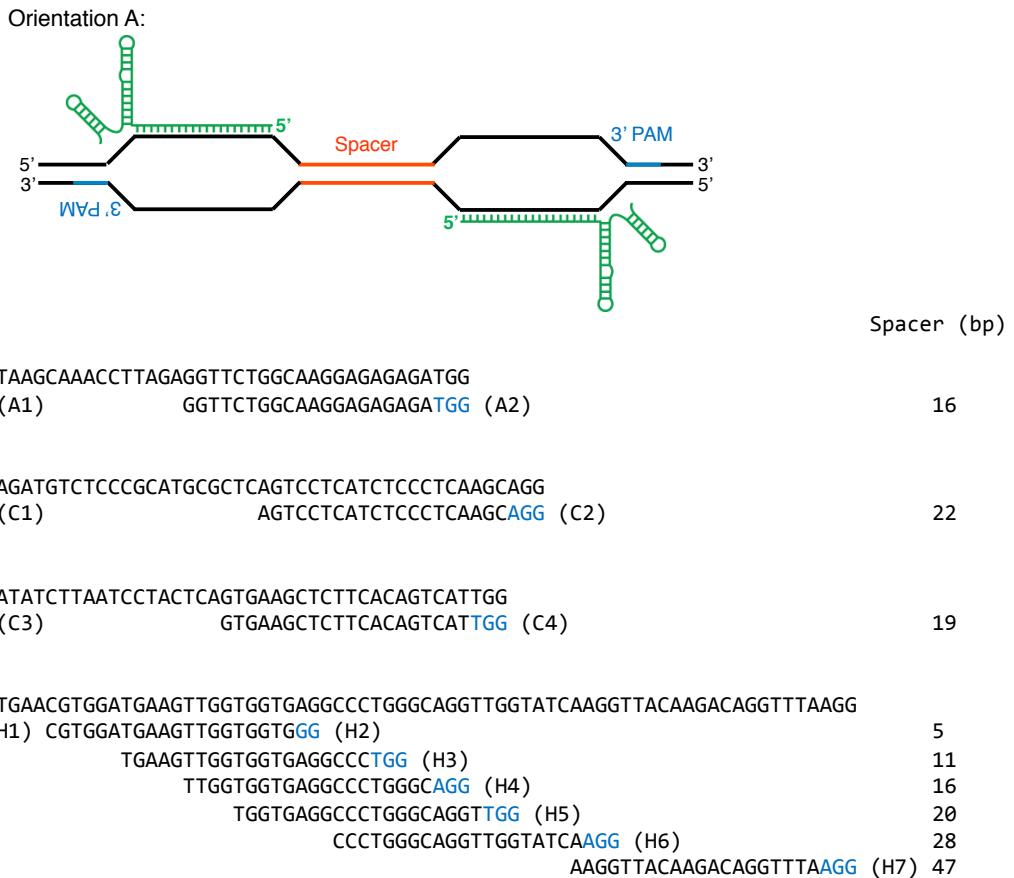
Supplementary Figure 3. Activities of *FokI-dCas9* fusion candidates combined with sgRNA pairs of different orientations and varying spacer lengths. Activity of *FokI-dCas9* fusion orientations in GFP disruption assay. The fusion architectures described in Figure 1b were tested for functionality by flow cytometry using the GFP loss-of-function reporter across all (a) orientation A sgRNA spacers and (b) orientation B sgRNA spacers (Figure 1c and Supplemental Figure 1). All *FokI-dCas9* fusion data shown are the results of single trials. Wild-type Cas9 and Cas9 nickase data are the average of two replicates, while the ‘no treatment’ negative control data is the average of 6 replicates, with error bars representing one standard deviation. The gray dotted line across the Y-axis corresponds to the average of the ‘no treatment’ controls performed on the same day.

a

Name	NLS-linker-Fok1	Fok1-linker-dCas9
<i>FokI</i> -(GGS)x3	GGS	GGSGGGSGGS
<i>FokI</i> -(GGS)x6	GGS	GGSGGGSGGSGGSGGS
<i>FokI</i> -L0	GGS	-
<i>FokI</i> -L1	GGS	MKIEQLPSA
<i>FokI</i> -L2	GGS	VRHKLKRVGS
<i>FokI</i> -L3	GGS	VPFLEPDNINGKTC
<i>FokI</i> -L4	GGS	GHGTGSTGS
<i>FokI</i> -L5	GGS	MSRPDPA
<i>FokI</i> -L6	GGS	GSAGSAAGSGEF
<i>FokI</i> -L7	GGS	SGSETPGTSESA
<i>FokI</i> -L8	GGS	SGSETPGTSESATPES
<i>FokI</i> -L9	GGS	SGSETPGTSESATPEGGS
NLS-(GGS)	GGS	GGSM
NLS-(GGS)x3	GGSGGGSGGS	GGSM
NLS-L1	VPFLEPDNINGKTC	GGSM
NLS-L2	GSAGSAAGSGEF	GGSM
NLS-L3	SIVAQLSRPDPA	GGSM
wild-type Cas9	N/A	N/A
Cas9 nickase	N/A	N/A

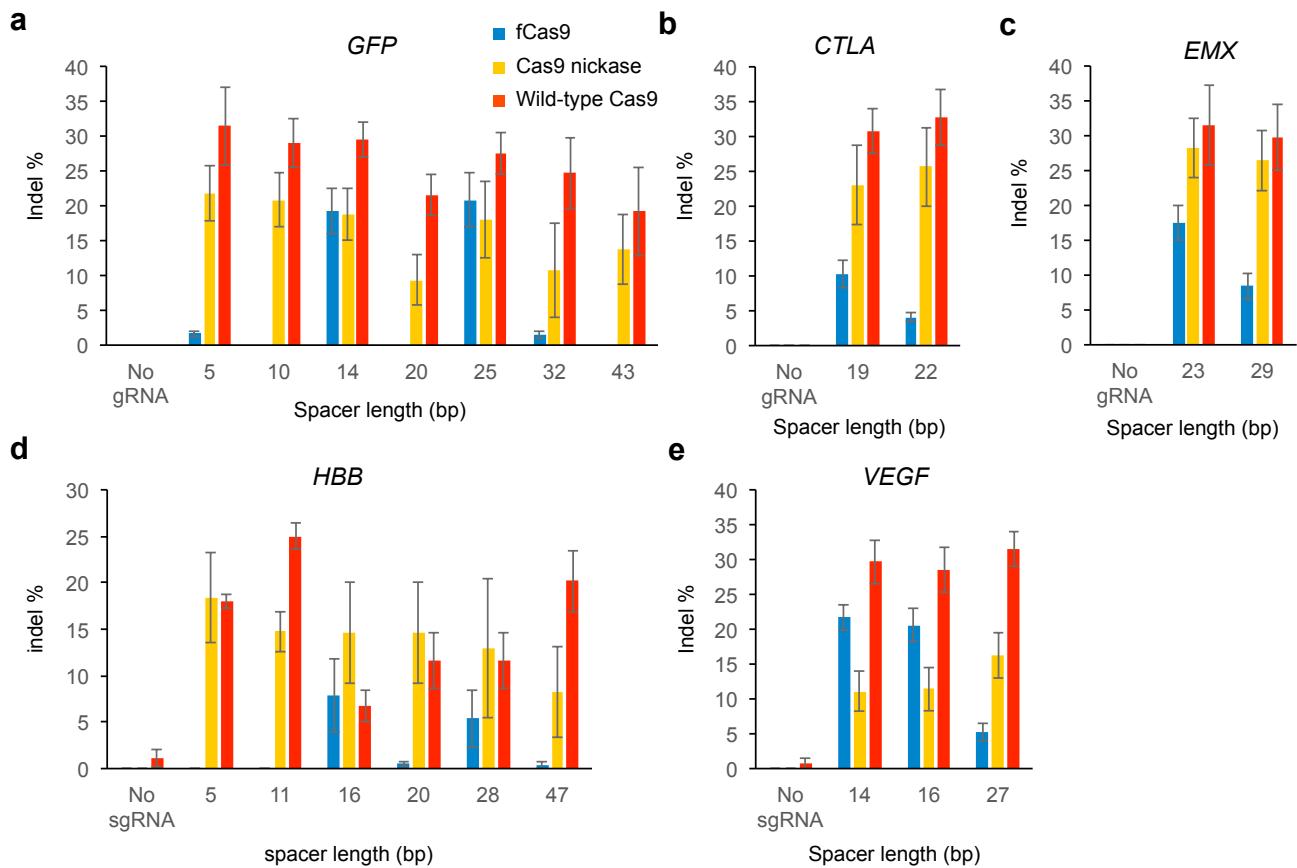
b

Supplementary Figure 4. Optimization of protein linkers in NLS-*FokI*-dCas9. (a) Table of all linker variants tested. Wild-type Cas9 and Cas9 nickase were included for comparison. The initial active construct NLS-*FokI*-dCas9 with a (GGS)₃ linker between *FokI* and dCas9 was tested across a range of alternate linkers. The final choice of linkers for fCas9 is highlighted in blue. (b) The activity of *FokI*-dCas9 fusions with linker variants. Each variant was tested across a range of spacer lengths from 5 to 43 bp using sgRNA pair orientation A. A control lacking sgRNA (grey) was included for each separate fusion construct. NLS-*FokI*-dCas9 variant L8 showed the best activity, approaching the activity of Cas9 nickase. Variants L4 through L9 show peak activity with 14- and 25-bp spacer lengths, suggesting two optimal spacer lengths roughly one helical turn of dsDNA apart.

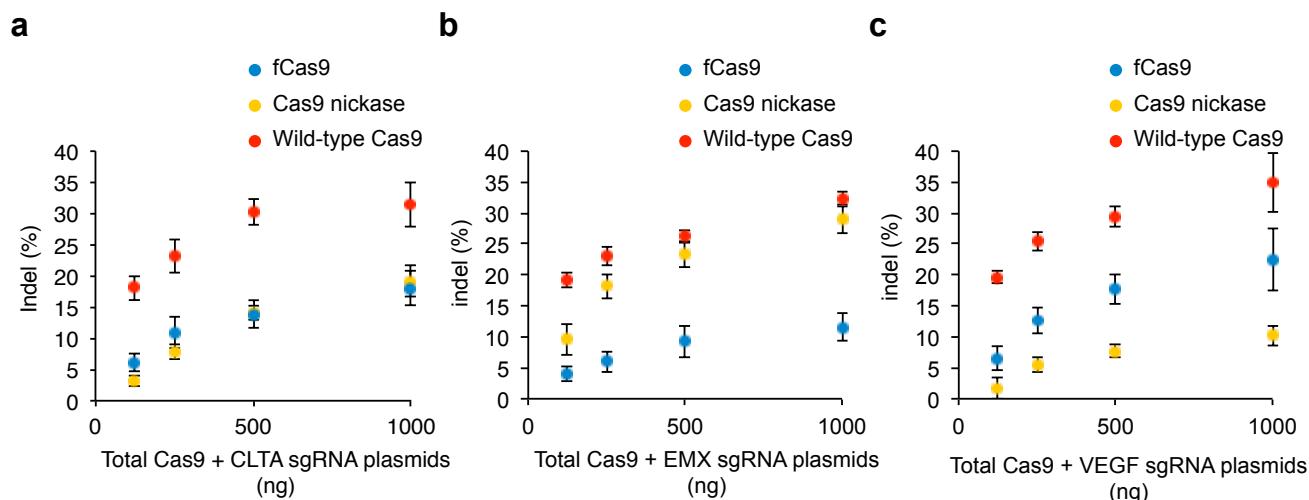


EMX		
CCCTTCTTCTGCTCGGACTCAGGCCCTCCTCCAGCTCTGCCGTTGACTTTGCTCCGGTTCTGG CCCCTTCTTCTGCTCGGACTC (E1)	GCCGTTGACTTTGCTCCGG (E2)	23
	TGTACTTTGCTCCGGTTCTGG (E3)	29
VEGF		
CCAGGAGCAAACCCCCCACCCCTTCAAAGCCCATTCCCTTTAGCCAGAGCCGGGGTGTGCAGACGG CCAAGGAGCAAACCCCCCACCC (V1)	ATTCCTCTTAGCCAGAGCCGG (V2)	14
	TCCCTCTTAGCCAGAGCCGGGG (V3)	16
	CCAGAGCCGGGGTGTGCAGACGG (V4)	27

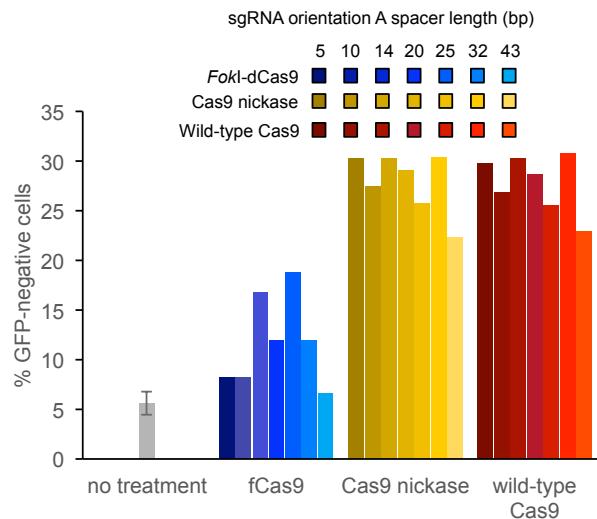
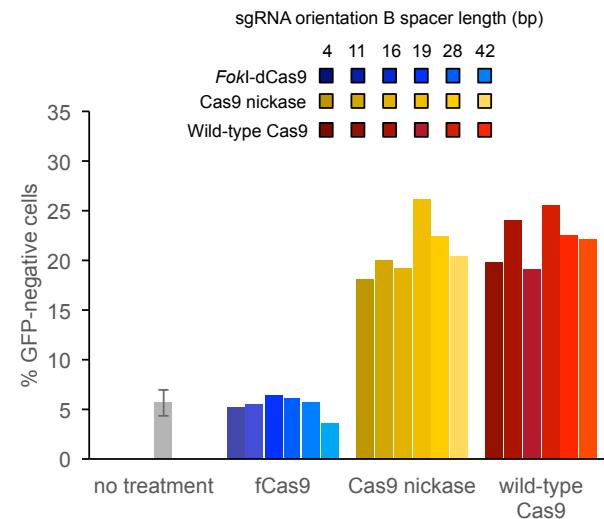
Supplementary Figure 5. Target DNA sequences in endogenous human *AAVS1*, *CLTA*, *HBB*, *EMX*, and *VEGF* genes. sgRNA target sites tested within endogenous human *AAVS1*, *CLTA*, *HBB*, *EMX*, and *VEGF* genes. Fourteen paired sgRNA target sites were chosen to test the activity of the optimized fCas9 fusion in an orientation in which the PAM is distal from the cleaved spacer sequence (orientation A). Together, these 14 sgRNA pairs enabled testing of fCas9 fusion variants across twelve spacer lengths ranging from 5 to 47 bp.



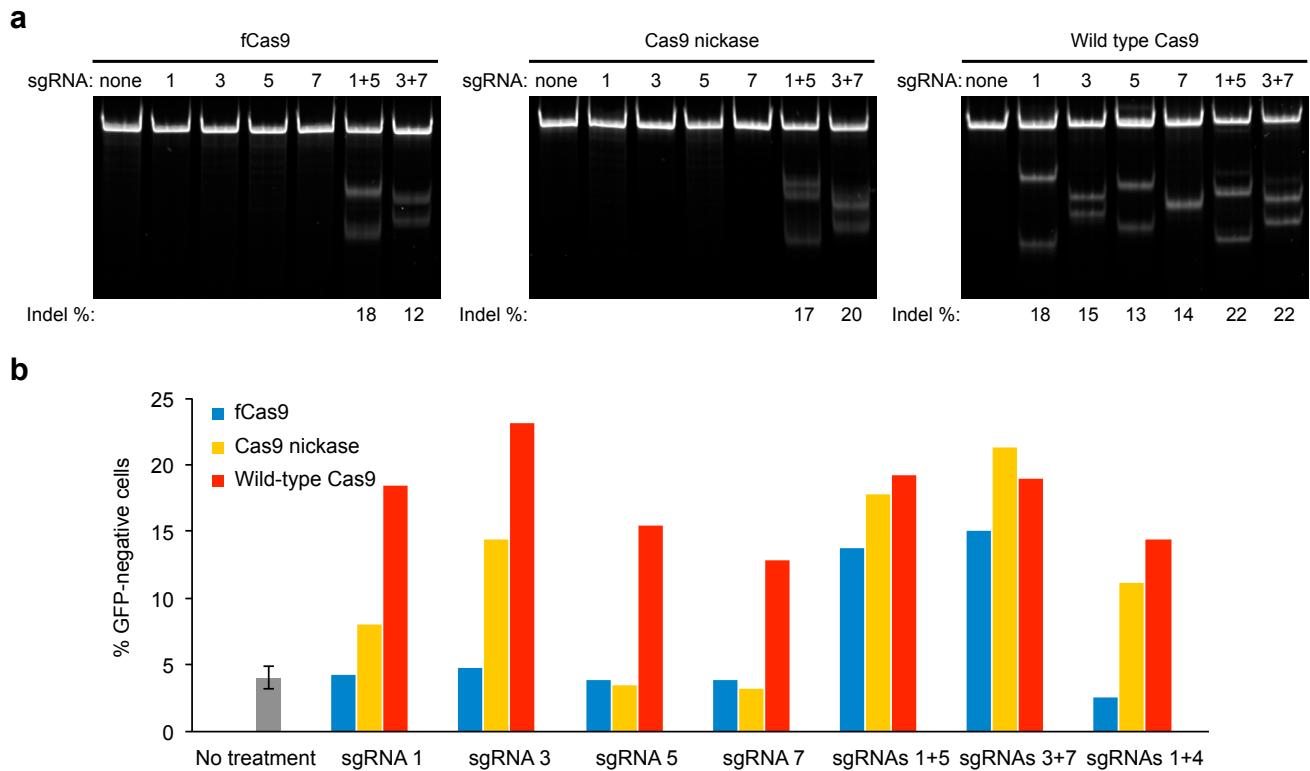
Supplementary Figure 6. Spacer length preference of genomic DNA modification by fCas9, Cas9 nickase, and wild-type Cas9. Indel modification efficiency for (a) pairs of sgRNAs targeting the *GFP* site, (b) pairs of sgRNAs targeting the *CTLA* site, (c) pairs of sgRNAs targeting the *EMX* site (d) pairs of sgRNAs targeting the *HBB* site, and (e) pairs of sgRNAs targeting the *VEGF* site. Error bars reflect standard error of the mean from three biological replicates performed on different days.



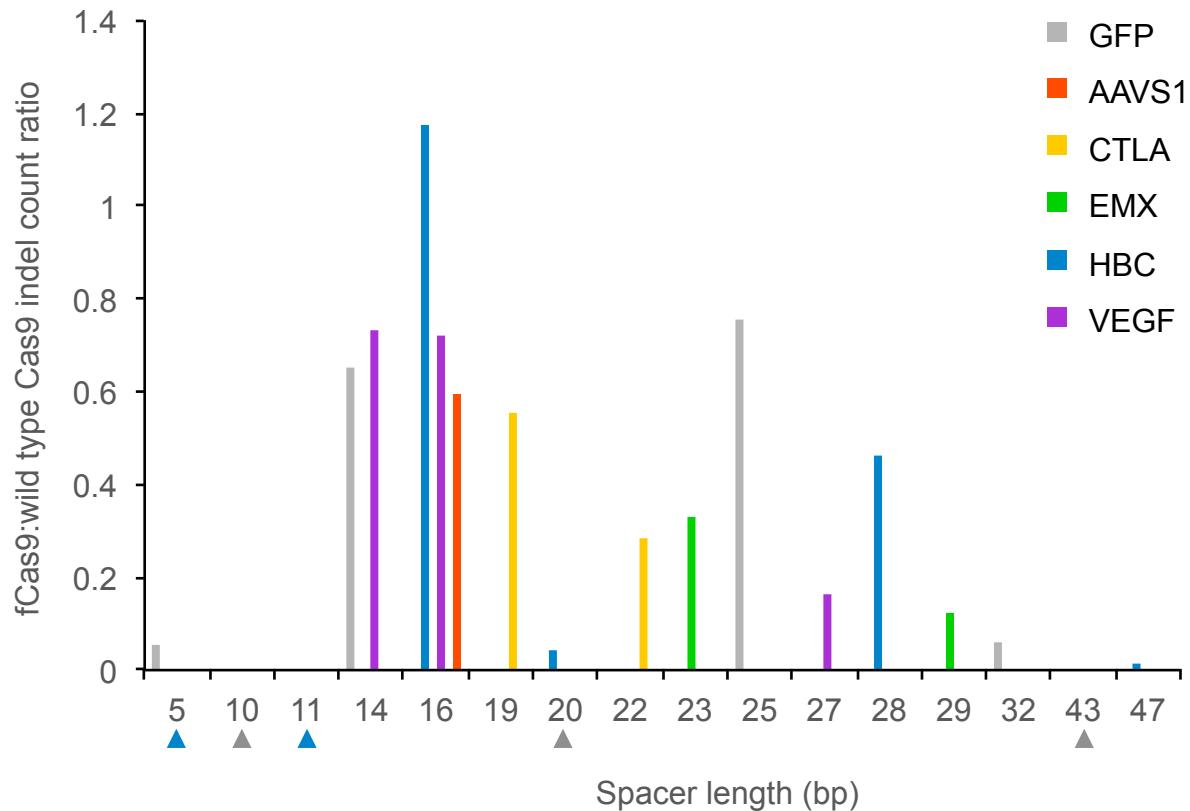
Supplementary Figure 7. Efficiency of genomic DNA modification by fCas9, Cas9 nickase, and wild-type Cas9 with varying amounts of Cas9 and sgRNA expression plasmids. Indel modification efficiency from a Surveyor assay of renatured target-site DNA amplified from a population of cells treated with fCas9, Cas9 nickase, or wild-type Cas9 and two target site sgRNAs. Either 700 ng of Cas9 expression plasmid with 250 ng of sgRNA expression plasmid (950 ng total), 350 ng of Cas9 expression plasmid with 125 ng of sgRNA expression plasmid (475 ng in total), 175 ng of Cas9 expression plasmid with 62.5 ng of sgRNA expression plasmid (238 ng in total) or 88 ng of Cas9 expression plasmid with 31 ng of sgRNA expression plasmid (119 ng in total) were transfected with an appropriate amount of inert, carrier plasmid to ensure uniform transfection of 950 ng of plasmid across all treatments. Indel modification efficiency for (a) sgRNAs spaced 19-bp apart targeting the *CLTA* site, (b) sgRNAs spaced 23 bp apart targeting the *EMX* site, and (c) sgRNAs spaced 14 bp apart targeting the *VEGF* site. Error bars represent the standard error of the mean from three biological replicates performed on separate days.

a**b**

Supplementary Figure 8. Dependence of fCas9 on sgRNA pair orientation. (a) GFP gene disruption by wild-type Cas9, Cas9 nickase, fCas9 using sgRNA pairs in orientation A. High activity of fCas9 requires spacer lengths of ~15 or 25 bp. (b) GFP gene disruption using sgRNA pairs in orientation B. Cas9 nickase, but not fCas9, accepts either orientation of sgRNA pairs. The “no treatment” control refers to cells receiving no plasmid DNA.



Supplementary Figure 9. Ability of fCas9, Cas9 nickase, and wild-type Cas9 to modify genomic DNA in the presence of a single sgRNA. (a) Surveyor assay of a genomic GFP target from DNA of cells treated with the indicated combination of Cas9 protein and sgRNA(s). Single sgRNAs do not induce genome modification at a detectable level (< 2% modification) for both fCas9 and Cas9 nickase. Wild-type Cas9 effectively modifies the GFP target for all tested single and paired sgRNAs. For both fCas9 and Cas9 nickase, appropriately paired sgRNAs induce genome modification at levels comparable to those of wild-type Cas9. (b) GFP gene expression loss by fCas9, but not Cas9 nickase or wild-type Cas9, depends on the presence of two sgRNAs. Four single sgRNAs were tested along with three sgRNA pairs of varying spacer length. In the presence of sgRNA pairs in orientation A with spacer lengths of 14 or 25 bp (sgRNAs 1+5, and sgRNAs 3+7, respectively), fCas9 is active, but not when a sgRNA pair with a 10-bp spacer (sgRNAs 1+4) is used.



Supplementary Figure 10. fCas9 indel frequency of genomic targets reflects sgRNA pair spacer length preference. The graph shows the relationship between spacer length (number of bp between two sgRNAs) and the indel modification efficiency of fCas9 normalized to the indel modification efficiency of the same sgRNAs co-expressed with wild-type Cas9 nuclease. Colored triangles below the X-axis denote spacer lengths that were tested but which yielded no detectable indels for the indicated target gene. These results suggest that fCas9 requires ~15 bp or ~25 bp between half-sites to efficiently cleave DNA.

a

Wild-type Cas9 nuclease modifications of VEGF on-target site:

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4747 gctgtttggaggcagaaatagggggtCCAGGAGCAAACCCCCCACCCtttccaaagcccATTCCCTTTAGCCAGAGCCGggtgtgcagacggcagtc (ref)
4577 gctgtttggaggcagaaatagggggtccagga-----agccggggtgtgcagacggcagtc
 58 gctgtttggaggcagaaatagggggtccaggagcaaactccccccaccccttccaaagcccattcccttttagc-----cggggtgtgcagacggcagtc
 54 gctgtttggaggcagaaatagggggtccaggag-----agccggggtgtgcagacggcagtc
 43 gctgtttggaggcagaaatag-----ccggggtgtgcagacggcagtc
33 gctgtttggaggcagaaatagggggtccaggag-----cggggtgtgcagacggcagtc
23 gctgtttggaggcagaaatagggggtccagg-----ccggggtgtgcagacggcagtc
22 gctgtttggaggcagaaatagggggtccaggagcaaactccccccaccccttccaaagcccattcccttttagccag-----ggtgtgcagacggcagtc
18 gctgtttggaggcagaaatagggggtccagg-----t-----agccggggtgtgcagacggcagtc

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Cas9 nickase modifications of VEGF on-target site:

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8770 gctgtttggaggcagaaatagggggtCCAGGAGCAAACCCCCCACCCtttccaaagcccATTCCCTTTAGCCAGAGCCGggtgtgcagacggcagtc (ref)
 78 gctgtttggaggcagaaatagggggtccag-----acggcagtc
 60 gctgtttggaggcagaaatagggggtccaggagcaaactccccccaccccttccaaagcc-----ggggtgtgcagacggcagtc
 58 gctgtttggaggcagaaatagggggtcca-----aagcccattcccttttagccagacggcggggtgtgcagacggcagtc
 56 gctgtttggaggcagaaataggggt-----gtgcagacggcagtc
 49 gctgtttggaggcagaaatagggggtccag-----ccggggtgtgcagacggcagtc
 37 gctgtttggaggcagaaatagggggtccagg-----gtgtgcagacggcagtc
 36 gctgtttggaggcagaaatagggggtccaggag-----cggggtgtgcagacggcagtc
 27 gctgtttggaggcagaaatag-----ccggggtgtgcagacggcagtc

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fCas9 nuclease modifications of VEGF on-target site:

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8959 gctgtttggaggcagaaatagggggtCCAGGAGCAAACCCCCCACCCtttccaaagcccATTCCCTTTAGCCAGAGCCGggtgtgcagacggcagtc (ref)
125 gctgtttggaggcagaaatagggggtccaggagcaaactccccca-----agccattcccttttagccagacggcggggtgtgcagacggcagtc
121 gctgtttggaggcagaaatagggggtccaggagcaaactccccccacccct-----ttcccttttagccagacggcggggtgtgcagacggcagtc
 77 gctgtttggaggcagaaatagggggtccaggagcaaactccccccacccct-----ttcccttttagccagacggcggggtgtgcagacggcagtc
 73 gctgtttggaggcagaaatagggggtccaggagcaaactccccca-----gcccattcccttttagccagacggcggggtgtgcagacggcagtc
 48 gctgtttggaggcagaaatagggggtccaggagcaaactccccccacccct-----attcccttttagccagacggcggggtgtgcagacggcagtc
 44 gctgtttggaggcagaaatagggggtccaggagcaaactccccccacccct-----agccagacggcggggtgtgcagacggcagtc
 24 gctgtttggaggcagaaatagggggtccaggagcaaactccccccacccctt-----aaagcccattcccttttagccagacggcggggtgtgcagacggcagtc
 22 gctgtttggaggcagaaatagggggtccaggagcaaactcccc-----aagcccattcccttttagccagacggcggggtgtgcagacggcagtc

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b

Wild-type Cas9 nuclease modifications of VEG_Off1:

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79248 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccCCAGGAGCAAACCCCtCCATCCCacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt (ref)
 800 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----caaaactccctccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 239 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccccc-----tcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
155 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccag-----caaaactccctccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 90 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccccc-----caaaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 71 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccccc-----tccctccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 54 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccccc-----tccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 53 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagga-----tccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 47 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccccc-----tccctccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt

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Cas9 nickase modifications of VEG_Off1:

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302573 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccCCAGGAGCAAACCCCtCCATCCCacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt (ref)
 28 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----atccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 13 cattcaacagatacttaactgaatgtctcagacaggacattctgac-----tccctccatcccacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 11 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccag-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
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 1 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
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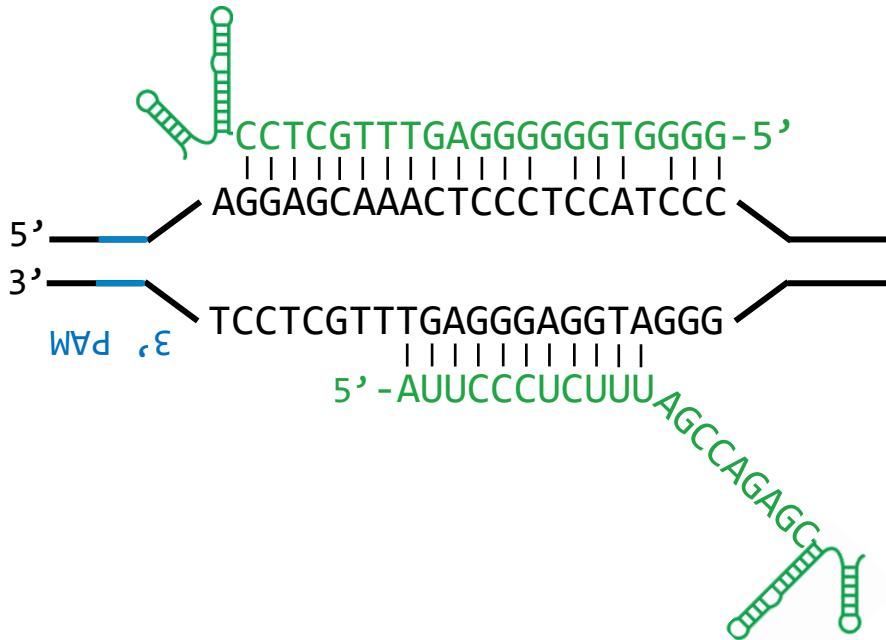
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fCas9 nuclease modifications of VEG_Off1:

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233567 cattcaacagatacttaactgaatgtctcagacaggacattctgacaccCCAGGAGCAAACCCCtCCATCCCacaatccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt (ref)
 6 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----gtcccttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 5 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 4 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 3 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 3 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
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 1 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----tccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt
 1 cattcaacagatacttaactgaatgtctcagacaggacattctgacacccccagg-----atccgtcttagatgtgcacacccaaacctcttaagaaaatagaaggatgt

```

c**d**

CCAGGAGCAA**ACT**CCCTCCATCCCACAAATCCGTCTTAGATGTGCACACCCAACCTCTAAGAAATA**GAGGATGATGGCCAGGTGCAGT**GATTCA**TGCCTG**
CCAGGAGCAAACT**CCC**C**CC**C**CC** (V1) **CCAGGAGCAa**act**CC**C**cc**a**C**cc****

Supplementary Figure 11. Modifications induced by Cas9 nuclease, Cas9 nickases, or fCas9 nucleases at endogenous loci. (a) Examples of modified sequences at the *VEGF* on-target site with wild-type Cas9 nuclease, Cas9 nickases, or fCas9 nucleases and a single plasmid expressing two sgRNAs targeting the *VEGF* on-target site (sgRNA V1 and sgRNA V2). For each example shown, the unmodified genomic site is the first sequence, followed by the top eight sequences containing deletions. The numbers before each sequence indicate sequencing counts. The sgRNA target sites are bold and capitalized. (b) Identical analysis as in (a) for *VEGF* off-target site 1VEG_Off1. (c) Potential binding mode of two sgRNAs to *VEGF* off-target site 1. The top strand is bound in a canonical mode, while the bottom strand binds the second sgRNA, sgRNA V2, through sgRNA:DNA base pairing that includes G:U base pairs. (d) Alternative off-target binding mode of two sgRNAs to *VEGF* off-target site 1. Binding of V1 sgRNA with a potential seed+PAM binding site 59 bases from the *VEGF* off-target site 1 that is closely related to V1 sgRNA target site. PAM shown in blue, mismatched positions shown in lower case red.

a

Spacer length (b)	Number of paired sgRNA sites in orientation A	Number of paired sgRNA sites in orientation B
-8	6874293	NC
-7	6785996	NC
-6	6984064	NC
-5	7023260	NC
-4	6487302	NC
-3	6401348	NC
-2	6981383	NC
-1	7230098	NC
0	7055143	NC
1	6598582	NC
2	6877046	NC
3	6971447	NC
4	6505614	5542549
5	6098107	5663458
6	6254974	6819289
7	6680118	6061225
8	7687598	5702252
9	6755736	7306646
10	6544849	6387485
11	6918186	6172852
12	6241723	5799496
13	6233385	7092283
14	6298717	7882433
15	6181422	7472725
16	6266909	6294684
17	6647352	6825904
18	6103603	6973590
19	5896092	6349456
20	6000683	5835825
21	5858015	6056352
22	6116108	6531913
23	5991254	6941816
24	6114969	6572849
25	6135119	5671641

b

Cas9 variant	Preferred spacer lengths (bp)	Total sites
fCas9	13 to 19, or 22 to 29, in orientation A	92354891
Cas9 nickase	-8 to 100 in orientation A 4 to 42 in orientation B	953048977

Supplementary Table 1. Paired sgRNA target site abundances for fCas9 and Cas9 nickase in the human genome. (a) Column 2 shows the number of sites in the human genome with paired sgRNA binding sites in orientation A allowing for a spacer length from -8 bp to 25 bp (column 1) between the

two sgRNA binding sites. sgRNA binding sites in orientation A have the NGG PAM sequences distal from the spacer sequence (CCNN₂₀-spacer-N₂₀NN). Column 3 shows the number of sites in the human genome with paired sgRNA binding sites in orientation B allowing for a spacer length from 4 to 25 bp (column 1) between the two sgRNA binding sites. sgRNA binding sites in orientation B have the NGG PAM sequences adjacent to the spacer sequence (N₂₀NNG spacer CCNN₂₀) . NC indicates the number of sites in the human genome was not calculated. Negative spacer lengths refer to target sgRNA binding sites that overlap by the indicated number of base pairs. **(b)** Sum of the number of paired sgRNA binding sites in orientation A with spacer lengths of 13 to 19 bp, or 22 to 29 bp, the spacer preference of fCas9 (**Supplementary Figure 8**). Sum of the number of paired sgRNA binding sites with spacer lengths of -8 bp to 100 bp in orientation A, or 4 to 42 bp in orientation B, the spacer preference of Cas9 nickases (4 to 42 bp in orientation B is based on **Figure 3b, c**, and -8 bp to 100 bp in orientation A is based on previous reports^{2,3}).

	Genomic target site
EMX_On	GAGTCCGAGCAGAAGAAGAA GGG
EMX_Off1	GAGgCCGAGCAGAAGAA ag A CGG
EMX_Off2	GAGTCCTAGCAGgAGAAGAA Ga G
EMX_Off3	GAGTC ta AGCAGAAGAAGAA Ga G
EMX_Off4	GAGT ta GAGCAGAAGAAGAA AGG
VEG_On	GGGTGGGGGGAGTTGCTCC TGG
VEG_Off1	GGaTGGaGGGAGTTGCTCC TGG
VEG_Off2	GGG a GGG t GGAGTTGCTCC TGG
VEG_Off3	cGGgGGaGGGAGTTGCTCC TGG
VEG_Off4	GGG ga GGGG a AGTTTGCTCC TGG
CLT2_On	GCAGATGTAGTGTTCAC GGG
CLT2_Off1	aCAaATGTAGT a TTTCCAC GGG
CLT2_Off2	cCAGATGTAGT a TTcCCAC GGG
CLT2_Off3	c t AGATG a AGTG c TTCCAC TGG

Supplementary Table 2. Known off-target substrates of Cas9 target sites in *EMX*, *VEGF*, and *CLTA*. List of genomic on-target and off-target sites of the EMX, VEGF, and CLTA are shown with mutations from on-target in lower case and red. PAMs are shown in blue.

a

Nuclease type:	wt Cas9	wt Cas9	Cas9 nickase	fCas9	wt Cas9	Cas9 nickase	fCas9
sgRNA pair target:	<i>CLTA</i>	<i>CLTA</i>	<i>CLTA</i>	<i>CLTA</i>	<i>GFP</i>	<i>GFP</i>	<i>GFP</i>
Total expression plasmids (ng):	1000	125	1000	1000	1000	1000	1000
<u><i>CLTA</i> Sites</u>							
<u><i>CLT2</i> On</u>							
Indels	3528	1423	3400	575	3	13	5
Total	10000	10000	10000	10000	10000	10000	10000
Modified (%)	35.280	14.230	34.000	5.750	0.030	0.130	0.050
P-value	<1.0E-300	<1.0E-300	<1.0E-300	1.4E-163			
On:off specificity	1	1		1			
<u><i>CLT2</i> Off1</u>							
Indels	316	44	2	2	1	3	3
Total	60620	64755	71537	63079	93883	91306	82055
Modified (%)	0.521	0.068	0.003	0.003	<0.002	0.003	0.004
P-value	1.3E-126	2.1E-16					
On:off specificity	68	209		>2850			
<u><i>CLT2</i> Off2</u>							
Indels	11	5	3	1	1	1	2
Total	72596	51093	59632	35541	69114	64412	39978
Modified (%)	0.015	0.010	0.005	0.003	<0.002	<0.002	0.005
P-value	6.5E-03						
On:off specificity	2328	1454		>2850			
<u><i>CLT2</i> Off3</u>							
Indels	11	10	0	0	1	1	1
Total	52382	44212	54072	48668	55670	58707	54341
Modified (%)	0.021	0.023	<0.002	<0.002	<0.002	<0.002	<0.002
P-value	2.7E-03	3.5E-03					
On:off specificity	1680	629		>2850			

b

Nuclease type:	wt Cas9	wt Cas9	Cas9 nickase	fCas9	wt Cas9	Cas9 nickase	fCas9
sgRNA pair:	<i>EMX</i>	<i>EMX</i>	<i>EMX</i>	<i>EMX</i>	<i>GFP</i>	<i>GFP</i>	<i>GFP</i>
Total expression plasmids (ng):	1000	125	1000	1000	1000	1000	1000
<u>EMX Site</u>							
<u>EMX_On</u>							
Indels	5111	2683	2267	522	0	0	2
Total	10000	10000	10000	10000	10000	10000	10000
Modified (%)	51.110	26.830	22.670	5.220	<0.002	<0.002	0.020
P-value	<1.0E-300	<1.0E-300	<1.0E-300	1.0E-154			
On:off specificity	1	1	1	1			
<u>EMX_Off1</u>							
Indels	386	122	7	1	4	9	7
Total	109787	83420	124564	88424	102817	90020	96526
Modified (%)	0.352	0.146	0.006	<0.002	0.004	0.010	0.007
P-value	1.3E-103	2.8E-37					
On:off specificity	145	183	>11222	>2584			
<u>EMX_Off2</u>							
Indels	74	58	3	6	3	0	4
Total	98568	94108	105747	78871	81717	79469	79193
Modified (%)	0.075	0.062	0.003	0.008	0.004	<0.002	0.005
P-value	3.2E-16	1.4E-12					
On:off specificity	681	435	>11222	>2584			
<u>EMX_Off3</u>							
Indels	736	178	20	14	12	11	17
Total	72888	65139	82348	59593	74341	73408	75080
Modified (%)	1.010	0.273	0.024	0.023	0.016	0.015	0.023
P-value	2.5E-202	3.1E-44					
On:off specificity	51	98	>11222	>2584			
<u>EMX_Off4</u>							
Indels	4149	620	3	3	6	7	5
Total	107537	91695	91368	91605	111736	119643	128088
Modified (%)	3.858	0.676	0.003	0.003	0.005	0.006	0.004
P-value	<1.0E-300	1.9E-202					
On:off specificity	13	40	>11222	>2584			

C

Nuclease type:	wt Cas9	wt Cas9	Cas9 nickase	fCas9	wt Cas9	Cas9 nickase	fCas9
sgRNA pair:	VEGF	VEGF	VEGF	VEGF	GFP	GFP	GFP
Total expression plasmids (ng):	1000	125	1000	1000	1000	1000	1000
<u>VEGF Sites</u>							
<u>VEG_On</u>							
Indels	5253	2454	1230	1041	8	0	1
Total	10000	10000	10000	10000	10000	10000	10000
Modified (%)	52.530	24.540	12.300	10.410	0.080	<0.002	0.010
P-value	<1.0E-300	<1.0E-300	<1.0E-300	6.6E-286			
On:off specificity	1	1	1	1			
<u>VEG_Off1</u>							
Indels	2950	603	22	0	0	4	1
Total	82198	71163	90434	77557	74765	79738	74109
Modified (%)	3.589	0.847	0.024	<0.002	<0.002	0.005	<0.002
P-value	<1.0E-300	3.2E-188	2.5E-06				
On:off specificity	15	29	506	>5150			
<u>VEG_Off2</u>							
Indels	863	72	3	3	0	2	1
Total	102501	49836	119702	65107	54247	65753	61556
Modified (%)	0.842	0.144	0.003	0.005	<0.002	0.003	<0.002
P-value	3.5E-159	9.6E-24					
On:off specificity	62	170	>6090	>5150			
<u>VEG_Off3</u>							
Indels	260	33	3	2	3	1	0
Total	91277	83124	90063	84385	62126	68165	69811
Modified (%)	0.285	0.040	0.003	0.002	0.005	<0.002	<0.002
P-value	6.8E-54	1.0E-05					
On:off specificity	184	618	>6090	>5150			
<u>VEG_Off4</u>							
Indels	1305	149	3	2	3	2	4
Total	59827	41203	65964	57828	60906	61219	62162
Modified (%)	2.181	0.362	0.005	0.003	0.005	0.003	0.006
P-value	<1.0E-300	2.7E-54					
On:off specificity	24	68	>6090	>5150			

d

Nuclease type:	Cas9 nickase	fCas9	Cas9 nickase	fCas9
sgRNA pair:	VEGF	VEGF	GFP	GFP
Total expression plasmids (ng):	1000	1000	1000	1000
<u>VEGF Sites</u>				
<u>VEG_On</u>				
Indels	2717	2122	10	13
Total	10000	10000	10000	10000
Modified (%)	27.170	21.220	0.100	0.130
P-value	<1.0E-300	<1.0E-300		
On:off specificity	1	1		
<u>VEG_Off1</u>				
Indels	67	30	3	2
Total	302573	233567	204454	190240
Modified (%)	0.022	0.013		
P-value	5.9E-12	2.5E-06		
On:off specificity	1227	1652		

Supplementary Table 3. Cellular modification induced by wild-type Cas9, Cas9 nickase, and fCas9 at on-target and off-target genomic sites. (a) Results from sequencing *CLTA* on-target and previously reported genomic off-target sites amplified from 150 ng genomic DNA isolated from human cells treated with a plasmid expressing either wild-type Cas9, Cas9 nickase, or fCas9 and a single plasmid expressing two sgRNAs targeting the *CLTA* on-target site (sgRNA C3 and sgRNA C4). As a negative control, transfection and sequencing were performed as above, but using two sgRNAs targeting the *GFP* gene on-target site (sgRNA G1, G2 or G3 and sgRNA G4, G5, G6 or G7). Indels: the number of observed sequences containing insertions or deletions consistent with any of the three Cas9 nuclease-induced cleavage. Total: total number of sequence counts while only the first 10,000 sequences were analyzed for the on-target site sequences. Modified: number of indels divided by total number of sequences as percentages. Upper limits of potential modification were calculated for sites with no observed indels by assuming there is less than one indel then dividing by the total sequence count to arrive at an upper limit modification percentage, or taking the theoretical limit of detection (1/49,500), whichever value was larger. P-values: For wild-type Cas9 nuclease, Cas9 nickase or fCas9 nuclease, P-values were calculated as previously reported¹⁸ using a two-sided Fisher's exact test between each sample treated with two sgRNAs targeting the *CLTA* on-target site and the control sample treated with two sgRNAs targeting the *GFP* on-target site. P-values of < 0.0045 were considered significant and shown based on conservative multiple comparison correction using the Bonferroni method. On:off specificity is the ratio of on-target to off-target genomic modification frequency for each site. (b) Experimental and analytic methods as in (a) applied to *EMX* target sites using a single plasmid expressing two sgRNAs targeting the *EMX* on-target site (sgRNA E1 and sgRNA E2). (c) Experimental and analytic methods as in (a) applied to *VEGF* target sites using a single plasmid expressing two sgRNAs targeting the *VEGF* on-target site (sgRNA V1 and sgRNA v2). (d) Experimental and analytic methods as in (a) applied to *VEGF* on-target and *VEGF* off-target site 1 amplified from 600 ng genomic DNA to increase detection sensitivity to 1/198,000.

Genomic target site

SiteA_On	CCG <u>CCTCTCTGAGCCTCAGTTCTTATCCAATTGATCTAAAGGTGGAATGGACATGCTGG</u>
SiteA_Off1	CCG <u>CCTCTCTGAGCCTCAGTTCTCATCAGCTGG---gAAAGagGGgcTGGACAaGatGaTAT</u>
SiteA_Off2	CCA <u>CCTCTCTGAGCCTCAGTTCTCACCTGTAATTGGGAATTATAACTAACCTTCCA</u>
SiteA_Off3	CCA <u>CCTCTCTGAGCCTCAGTTCTTAATCTGAAAAAGAGATTAATAACTTCCCTCACT</u>
SiteB_On	CCA <u>CTGTGCCTGGCCTCTAATTCTAATTCTAGAATCATAAGAAGTTGCAAACATTGTACAGTGGGTGG</u>
SiteB_Off1	CCA <u>CTGTGCCTGGCCTCTAATTcTTCATTAAGGTGACATAGTCATTCTGGAGCTTAAGCTGCTTCTG</u>
SiteB_Off2	CCA <u>CTGTGCCTGGCCTCTAATTgCTTTTTTATAAAATTCTTTGGATTTCTTAGTCATGTTATCTTGAA</u>
SiteB_Off3	CCA <u>CTGTGCCTGGCCTCTAATTgTGCTGTTTATAAGTTGGACTGGCATGGCCTACTTAAATTTCAGG</u>
SiteB_Off4	CCA <u>CTGTGCCTGGCCTCTAATTAAAGTCTTACATTTCATCTGTCAGACTGTTGACCCAGCTTGG</u>
SiteC_On	CCT <u>CCGTGCCTGGCCTCAAATGCGTGCATTTGATTGACTGAATCCACTGCTTCCTCACAAACTGGG</u>
SiteC_Off1	CCT <u>CCGTGCCTGGCCTCAAATGCTTGTATTGTCACAGATGTTGTAATAGGTTGCCAGTTGGC</u>
SiteC_Off2	CCA <u>CCGTGCCTGGCCTCAAATGCAGACCTGTTTTGGAAAAGCTTACTGAATAAGGAAGGAAAAGAT</u>
SiteC_Off3	CCA <u>CCGTGCCTGGCCTCAAATGCATTTCTATACAGGTCCAGTCAGAGTAGTGGCCAGTCCCACTCATG</u>
SiteC_Off4	CCA <u>CCGTGCCTGGCCTCAAATGgCTTTAAACATAAGAAAATGTGCTCAGCCTCACTCATATAAAAGT</u>
SiteC_Off5	CCA <u>CCGTGCCTGGCCTCAAATGtCTTTCTATGTCAGAAAATTCAATCCTATGAGCTGGCTATAATT</u>

Supplementary Table 4. Genomic fCas9 target sites A, B and C with highly similar off-target substrates. List of genomic on-target and off-target sites are shown with mutations from on-target in lower case and red. PAMs are shown in blue. Site A corresponds to the human genomic locus chr1:21,655,401-21,655,461. Site B corresponds to the human genomic locus chr2:31,485,447-31,485,516. Site C corresponds to the human genomic locus chr3:48,747,484-48,747,552

a

		fCas9	Cas9 nickase
Nuclease type:	none		
gRNA pair target:	none	SiteA	SiteA
A Sites			
SiteA_On			
Indels	0	1217	4814
Total	36079	79671	73731
Modified (%)	<0.001	1.528	6.529
P-value		1.3E-196	<1.0E-300
On:off specificity	1	1	1
SiteA_Off1			
Indels	3	4	42
Total	62539	87323	96207
Modified (%)	0.005	0.005	0.044
P-value			8.9E-07
On:off specificity		>333	150
SiteA_Off2			
Indels	3	3	147
Total	72314	80940	115745
Modified (%)	0.004	0.004	0.127
P-value			4.2E-27
On:off specificity		>412	51
SiteA_Off3			
Indels	4	4	85
Total	76285	103657	108214
Modified (%)	0.005	0.004	0.079
P-value			2.1E-15
On:off specificity		>396	83

b

		fCas9	Cas9 nickase
Nuclease type:	none		
gRNA pair target:	none	SiteB	SiteB
B Sites			
SiteB_On			
Indels	1	1333	1054
Total	100164	82939	10000
Modified (%)	<0.001	1.607	10.540
P-value		<1.0E-300	<1.0E-300
On:off specificity		1	1
SiteB_Off1			
Indels	0	7	8
Total	81400	75845	91087
Modified (%)	<0.001	0.009	0.009
P-value			
On:off specificity		>174	>1200
SiteB_Off2			
Indels	1	2	27
Total	92707	100170	165464
Modified (%)	0.001	0.002	0.016
P-value			9.4E-05
On:off specificity		>805	646
SiteB_Off3			
Indels	0	2	14
Total	91872	102947	117175
Modified (%)	<0.001	0.002	0.012
P-value			4.9E-04
On:off specificity		>827	882

c

		fCas9	Cas9 nickase
Nuclease type:	none		
gRNA pair target:	none	SiteB	SiteB
C Sites			
SiteC_On			
Indels	3	893	769
Total	85113	69013	10000
Modified (%)	0.004	1.294	7.690
P-value		<1.0E-300	<1.0E-300
On:off specificity		1	1

SiteC_Off1

Indels	0	5	329
Total	114777	95225	117836
Modified (%)	<0.001	0.005	0.279
P-value			8.6E-98
On:off specificity		>246	28

SiteC_Off2

Indels	1	2	20
Total	70502	73601	112343
Modified (%)	0.001	0.003	0.018
P-value			1.0E-03
On:off specificity		>476	432

SiteC_Off3

Indels	1	3	106
Total	43908	39215	52210
Modified (%)	0.002	0.008	0.203
P-value			6.0E-27
On:off specificity		>169	38

SiteC_Off4

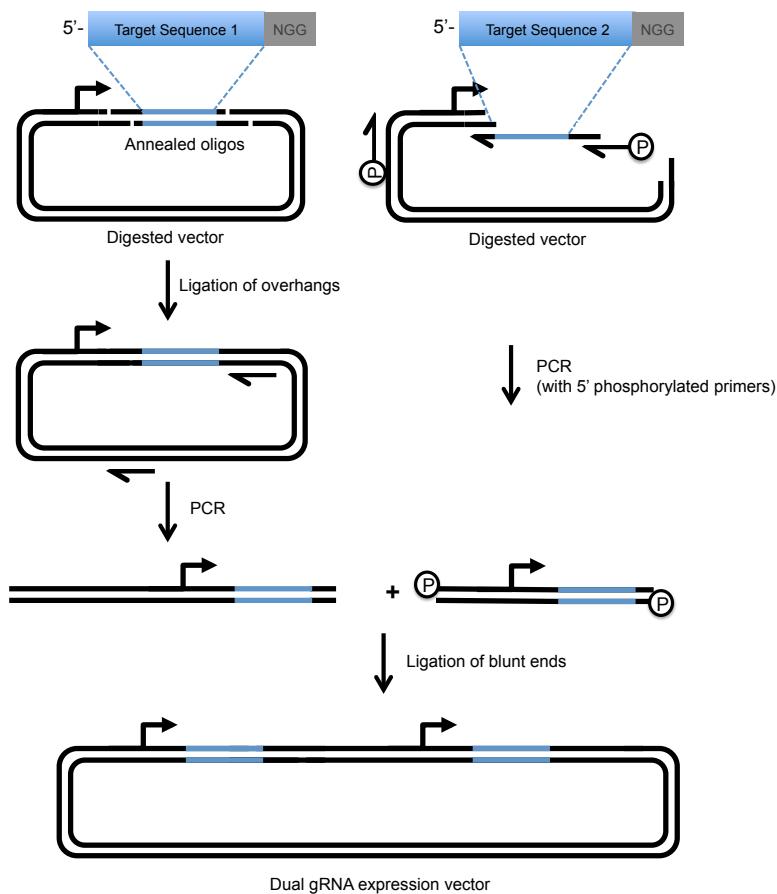
Indels	4	0	4
Total	143409	54192	70430
Modified (%)	0.003	<0.001	0.006
P-value			
On:off specificity		>1294	>1354

Supplementary Table 5. Cellular modification induced by Cas9 nickase and fCas9 at target sites A, B and C and highly similar off-target genomic sites. (a) Results from sequencing genomic site A on-target and highly similar genomic off-target sites amplified from 300 ng genomic DNA isolated from human cells treated with a plasmid expressing either Cas9 nickase, or fCas9 and two separate linear PCR fragments each expressing a distinct gRNA targeting genomic site A. As a negative control, transfection and sequencing were performed on cells with an inert plasmid. Indels: the number of observed sequences containing insertions or deletions consistent with any of the two Cas9 nuclease-induced cleavage. Total: total number of sequence counts while only the first 10,000 sequences were analyzed for some on-target site sequences. Modified: number of indels divided by total number of sequences as percentages. Upper limits of potential modification were calculated for sites with no observed indels by assuming there is less than one indel then dividing by the total sequence count to arrive at an upper limit modification percentage, or taking the theoretical limit of detection (1/99,000), whichever value was larger. P-values: For Cas9 nickase or fCas9 nuclease, P-values were calculated as previously reported¹⁸ using a two-sided Fisher's exact test between each sample treated with two gRNAs targeting the genomic site A on-target site and the control sample treated with no Cas9 or gRNAs. P-values of < 0.005 were considered significant and shown based on conservative multiple comparison correction using the Bonferroni method. On:off specificity is the

ratio of on-target to off-target genomic modification frequency for each site. **(b)** Experimental and analytic methods as in **(a)** applied to genomic site B. PCR amplification of SiteB_Off4 was not successful. **(c)** Experimental and analytic methods as in **(a)** applied to genomic site C. PCR amplification of SiteC_Off5 was not successful.

SUPPLEMENTARY NOTES

Construction of a single plasmid expressing two separate sgRNAs. The strategy used to construct each dual sgRNA expression plasmid is shown in the diagram below. A linear DNA fragment containing a Hu6 promoter (bent arrow), followed by the target site, followed by the sgRNA constant region was constructed by PCR amplification using digested expression vector and nested primers (with 5' phosphate modifications). The phosphorylated DNA fragments were joined by blunt-end ligation with a DNA fragment containing the vector backbone and a Hu6 promoter driving expression of the second sgRNA. Ultimately, two distinct gRNA constructs are expressed from separate (but identical) Hu6 promoters on the same plasmid construct.



DNA Sequence-Processing Algorithms. All scripts were written in bash. Scripts are available upon request.

Computational Search for Potential Target Sites

1) The Patmatch program⁴ was used to search the human genome (GRCh37/hg19 build) for pattern sequences corresponding to Cas9 binding sites (CCN N²⁰ spacer N²⁰NGG for Orientation A and

N^{20} NGG spacer CCN N^{20} for Orientation B)

Identification of Indels in Sequences of Genomic Sites

1) Sequence reads were initially filtered removing reads of less than 50 bases and removing reads with greater than 10% of the Illumina base scores not being B-J:

Example SeqA-1st read:

TTCTGAGGGCTGCTACCTGTACATCTGCACAAGATTGCCTTACTCCATGCCTTCTTCT
GCTCTAACTCTGACAATCTGTCTGCCATGCCATAAGCCCCTATTCTTCTGTAACCCCAAGATGGT
ATAAAAGCATCAATGATTGGGC

Example SeqA-2nd read:

AAAACCTAAAGAAATGCCAATCATTGATGCTTTATACCATCTGGGTTACAGAAAGAA
TAGGGGCTTATGGCATGGCAAGACAGATTGTCAGAGTTAGAGCAGAAGAAGAAAGGCATGGAGT
AAAGGCAATCTGTGCAGATGTACAGGTAA

2) Find the first 20 bases four bases from the start of the reverse complement of SeqA-2nd read in SeqA-1st read allowing for 1 mismatch:

Reverse complement of SeqA-2nd read:

TTACCTGTACATCTGCACAAGATTGCCTTACTCCATGCCTTCTTCTGCTCTA
ACTCTGACAATCTGTCTGCCATGCCATAAGCCCCTATTCTTCTGTAACCCCAAGATGGT
ATAAAAGCATCAATGATTGGGCATTCTTGAGTTT

Position in SeqA-1st read

TTCTGAGGGCTGCTACCTGTACATCTGCACAAGATTGCCTTACTCCATGCCTTCTTCTGCT
CTAACTCTGACAATCTGTCTGCCATGCCATAAGCCCCTATTCTTCTGTAACCCCAAGATGGTATA
AAAGCATCAATGATTGGGC

3) Align and then combine sequences, removing any sequence with greater than 5% mismatches in the simple base pair alignment:

Combination of SeqA-1st read and SeqA-2nd read:

TTCTGAGGGCTGCTACCTGTACATCTGCACAAGATTGCCTTACTCCATGCCTTCT
TCTTCTGCTCTAACTCTGACAATCTGTCTGCCATGCCATAAGCCCCTATTCTTCTGTAAC
CCCAAGATGGTATAAAAGCATCAATGATTGGCATTCTTGAGTTT

4) To identify the target site the flanking genomic sequences were searched for with the Patmatch program⁴ allowing for varying amounts of bases from 1 to 300 between the flanking genomic sequences:

Target Site	Downstream genomic sequence	Upstream genomic sequence
EMX_On	GGCCTGCTTCGTGGCAATGC	ACCTGGGCCAGGGAGGGAGG
EMX_Off1	CTCACTTAGACTTTCTCTCC	CTCGGAGTCTAGCTCTGCA
EMX_Off2	TGGCCCCAGTCTCTTCTA	CAGCCTCTGAACAGCTCCG
EMX_Off3	TGACTTGGCCTTGTAGGAA	GAGGCTACTGAAACATAAGT
EMX_Off4	TGCTACCTGTACATCTGCAC	CATCAATGATTGGCATTTC
VEG_On	ACTCCAGTCCCAAATATGTA	ACTAGGGGGCGCTCGGCCAC
VEG_Off1	CTGAGTCAACTGTAAGCATT	GGCCAGGTGAGTGATTCAT
VEG_Off2	TCGTGTCATCTGTTGTGC	GGCAGAGCCCAGCGGACACT
VEG_Off3	CAAGGTGAGCCTGGGTCTGT	ATCACTGCCAACAGAAGTGCA
VEG_Off4	TTGTAGGATGTTAGCAGCA	ACTTGCTCTTTAGAGAAC
CLT2_On	CTCAAGCAGGCCCGCTGGT	TTTGGACCAACCTTTG
CLT2_Off1	TGAGGTATTGTCATTGT	TAAGGGAGTATTACACCA
CLT2_Off2	TCAAGAGCAGAAAATGTGAC	CTTGCAGGGACCTTCTGATT
CLT2_Off3	TGTGTGAGGACTAACTCT	GATAGCAGTATGACCTGGG
SiteA_On	GCTCTGCCACCACAAAGCTTGGGCA	CCCTTGCAATCCATTCCCCCTACCA

SiteA_Off1	GGAGATGAACCAGCCTGCAGTCAG	ACTGATCTATGCCGTGCCCTTGTG
SiteA_Off2	CCCAGTCCCTATCACAAAAAAAGAT	ACATTGATCATCATGGCCACTGGAT
SiteA_Off3	TCCTGATGCCAGCACTCAGTGCCTG	AAGAGCACCAAGTACAGTCTGTGGC
SiteB_On	TTCCCAAACGTGCTGGATTACAGGC	TGCTACTGTGTAAGGGCATAGT
SiteB_Off1	CTCAGCCTCTCAAAGTGCTGGGATT	TATCTCCTCCCTTCCCTCCCTTC
SiteB_Off2	CTCCCCAAAGTGCTGGATTACAGGC	TTGGTTATAGAAACACCATTGAT
SiteB_Off3	CTCCCCAAAGTGCTGGATTACAAGG	GAATGTTAAGTTGTCCAGAGGCCA
SiteB_Off4	GCCTCCCAAAGTGCTGGGATTACAG	CCAGCACTTGGGAGGCCAAAGCGG
SiteC_On	CCTCAGCTTCCCAAACGTGCTGAGAT	GTGTGACCTTGCTTGGAACTGTG
SiteC_Off1	TCTCGACCTCCCTAACGTGCTGGGAT	CTTGCAGAAGAGTGCCAGTTGTGGT
SiteC_Off2	AATCTGCCCACCTCGGCCTCCAAA	TACCACTTTAAAATTCACCTCTC
SiteC_Off3	CTCCCCAAAGTGCTGGTATTACAGGT	CTTTGTCTTAATAATTCCCTATT
SiteC_Off4	CTGCCTCAGCCTCCGAAGTGCTAG	ATAATCCCAGCACTTGAAGGCTG

4) Any target site sequences corresponding to the same size as the reference genomic site in the human genome (GRCh37/hg19 build) were considered unmodified and any sequences not the reference size were considered potential insertions or deletions. Sequences not the reference size were aligned with ClustalW⁵ to the reference genomic site. Aligned sequences with more than one insertion or one deletion in the DNA spacer sequence in or between the two half-site sequences were considered indels. Since high-throughput sequencing can result in insertions or deletions of one base pairs (mis-phasing) at a low but relevant rates - indels of two bp are more likely to arise from Cas9 induced modifications.

Oligonucleotides Used in This Study

All oligonucleotides were purchased from Integrated DNA Technologies. '/5Phos/' indicates 5' phosphorylated oligonucleotides.

dCas9-NLS-FokI primers:

Cas9_Exp_CNF_Fok1+PI as-Fwd	CGGCGAGATAAACTTTAA TGACCGGTATCATCACCA
Cas9_Exp_CNF_Cas9coD 10-Rev	CCAACGGAATTAGTGCCGATAGCTAACCAATAGAATACTTTTATC
Cas9_Exp_CNF_Cas9coD 10-Fwd	GATAAAAAGTATTCTATTGGTTAGCTATGGCACTAATTCCGTTGG
Cas9_Exp_CNF_Cas9coH 850-Rev	TTCAAAAAGGATTGGGTACAATGGCATCGACGTCGTAATCAGATAAAC
Cas9_Exp_CNF_Cas9coH 850-Fwd	GTTTATCTGATTACGACGTCGATGCCATTGTACCCAATCCTTTGAA
Cas9_Exp_CNF_(Cas9)N LS+GGS-Fok-Rev	TTGGGATCCAGAACCTCCCTGCAGCCTGTCATCG
Cas9_Exp_CNF_(Cas9)N LS+GGS3-Fok-Rev	TTGGGATCCAGAACCTCC GCTGCCGCACTCCACCTGA
Cas9_Exp_CNF_(Cas9)N LS+GGS-Fok-Fwd	TCCTGCAGCCTGTCATCG
Cas9_Exp_CNF_(Cas9)N LS+GGS3-Fok-Fwd	CGATGACAAGGCTGCAGGAGGAGGTTCTGGATCCAA
Cas9_Exp_CNF_Fok1+PI as-Rev	CGATGACAAGGCTGCAGGA TCAGGTGGAAGTGGCGGCAGC GGAGGTTCTGGATCCAA
NLS-dCas9-FokI primers:	TGGTGATGATGACCGGTCA TTAAAAGTTATCTCGCCG
Cas9_Exp_NCF_Fok1+PI as-Fwd	CGGCGAGATAAACTTTAA TGACCGGTATCATCACCA

Cas9_Exp_NCF_Plasmid+F	TAGGGAGAGCCGCCACCATGGACTACAAAGACCATGACGG
LAG(NLS-Fok1-Rev	
Cas9_Exp_NCF_NLS	TAAACCAATAGAATACTTTTATC CATAGGTACCCCGCGGTGAATG
+Cas9coD10-Rev	
Cas9_Exp_NCF_Cas9coD	GATAAAAAGTATTCTATTGGTTAGCTATCGGCACTAATTCCGTTGG
10-Fwd	
Cas9_Exp_NCF_Cas9coH	TTCAAAAAGGATTGGGGTACAATGGCATCGACGTCGTAATCAGATAAAC
850-Rev	
Cas9_Exp_NCF_Cas9coH	GTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTGAA
850-Fwd	
Cas9_Exp_NCF_Cas9End	TTGGGATCCAGAACCTCCGTACCCCCAAGCTGTG
+GGS-Fok-Rev	TTGGGATCCAGAACCTCC GCTGCCGCACCCACCTGA
Cas9_Exp_NCF_Cas9End	GTCACCCCCAAGCTGTG
+GGS3-Fok-Rev	
Cas9_Exp_NCF_Cas9End	CACAGCTTGGGGGTGACGGAGGTTCTGGATCCAA
+GGS-Fok-Fwd	CACAGCTTGGGGTGAC TCAGGTGGAAGTGGCGGCAGC
Cas9_Exp_NCF_Cas9End	GGAGGTTCTGGATCCAA
+GGS3-Fok-Fwd	
Cas9_Exp_NCF_Fok1+Pl	TGGTGATGATGACCGGTCA TTAAAAGTTATCTCGCCG
asm-Rev	
FokI-dCas9-NLS primers:	
Cas9_Exp_FCN_Plasmid+F	TAGGGAGAGCCGCCACCATGGATCCAACTAGTCAAAAG
ok-Fwd	
Cas9_Exp_FCN_Fok1GG	ACCAATAGAATACTTTTATCCATGCTGCCACCAAAGTTATCTC
S+Cas-Rev	
Cas9_Exp_FCN_Fok1GG	ACCAATAGAATACTTTTATCCATGCTGCCGCACCCACCTG
S3+Cas-Rev	
Cas9_Exp_FCN_Cas9coD	GATAAAAAGTATTCTATTGGTTAGCTATCGGCACTAATTCCGTTGG
10-Fwd	
Cas9_Exp_FCN_Cas9coH	CCAACGGAATTAGTGCCGATAGCTAACCAATAGAATACCTTTATC
850-Rev	
Cas9_Exp_FCN_Cas9coH	GTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTGAA
850-Fwd	
Cas9_Exp_FCN_Plasmid+F	CTTTGACTAGTTGGATCCCATGGTGGCGGCTCTCCCTA
ok-Rev	
NLS-FokI-dCas9 primers:	
Cas9_Exp_NFC_Plasmid+F	TAGGGAGAGCCGCCACCATGGACTACAAAGACCATGACGG
LAG-Fwd	
Cas9_Exp_FCN_Fok1GG	ACCAATAGAATACTTTTATCCATGCTGCCACCAAAGTTATCTC
S+Cas-Rev	

Cas9_Exp_FCN_Fok1GG S3+Cas-Rev	ACCAATAGAATACTTTTATCCATGCTGCCGCCACTTCCACCTG
Cas9_Exp_FCN_Cas9coD 10-Fwd	GATAAAAAGTATTCTATTGGTTAGCTATCGGCACTAATTCCGTGG
Cas9_Exp_NFC_Cas9coH 850-Rev	CCAACGGAATTAGTGCCGATAGCTAACCAATAGAATACTTTTATC
Cas9_Exp_NFC_Cas9coH 850-Fwd	GTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTGAA
Cas9_Exp_NFC_Cas9End +PlasE-Rev	TGGTGATGATGACCGGTCA GTCACCCCCAAGCTGTG
Cas9_Exp_NFC_Cas9End +PlasE-Fwd	CACAGCTTGGGGGTGAC TGACCGGTATCATCACCA
Cas9_Exp_NFC_PlasS+F LAG-Rev	CCGTCATGGTCTTGTAGTCCATGGTGGCGGCTCTCCCTA
sgRNA_G1-top	ACACCCCTCGAACCTCACCTCGGG
sgRNA_G2-top	ACACCGTCGCCCTCGAACCTCACCTG
sgRNA_G3-top	ACACCCAGCTCGATCGGGTACCCAG
sgRNA_G4-top	ACACCGGTGAACCGCATCGAGCTGAG
sgRNA_G5-top	ACACCGCTGAAGGGCATCGACTTCAG
sgRNA_G6-top	ACACCGGCATCGACTCAAGGAGGAG
sgRNA_G7-top	ACACCCAAGGAGGACGGCAACATCCG
sgRNA_G8-top	ACACCACCATCTTCTCAAGGACGAG
sgRNA_G9-top	ACACCCAACTACAAGACCCGCGCCGG
sgRNA_G10-top	ACACCCCAGCCGAGGTGAAGTCGG
sgRNA_G11-top	ACACCGAAGTCGAGGGCGACACCG
sgRNA_G12-top	ACACCTTCGAACCTCACCTCGCGCG
sgRNA_G13-top	ACACCTCAGCTCGATCGGGTACCCG
sgRNA_G14-top	ACACCCGATGCCCTTCAGCTCGATGG
sgRNA_G1-bottom	AAAACCGCCGAGGTGAAGTCGAGGG
sgRNA_G2-bottom	AAAACAGGTGAAGTCGAGGGCGACG
sgRNA_G3-bottom	AAAACTGGTGAACCGCATCGAGCTGG
sgRNA_G4-bottom	AAAACTCAGCTCGATCGGGTACCCG
sgRNA_G5-bottom	AAAACTGAAGTCGATGCCCTTCAGCG
sgRNA_G6-bottom	AAAACTCCTCCTGAAGTCGATGCCG
sgRNA_G7-bottom	AAAACGGATTTGCCGTCCTCCTGG
sgRNA_G8-bottom	AAAACTCGTCCTGAAGAAGATGGTG
sgRNA_G9-bottom	AAAACCGGCGCGGGTCTTAGTTGG
sgRNA_G10-bottom	AAAACCGAACTTCACCTCGCGCGGG
sgRNA_G11-bottom	AAAACGGGTGTCGCCCTCGAACCTCG
sgRNA_G12-bottom	AAAACGCGCCGAGGTGAAGTCGAAG
sgRNA_G13-bottom	AAAACGGTGAACCGCATCGAGCTGAG
sgRNA_G14-bottom	AAAACCATCGAGCTGAAGGGCATCGG

sgRNA_A1-top	AAAACGGATCCTCTGGCTCCATCG
sgRNA_C1-top	ACACCTGGCCTGCTTAGACTTGG
sgRNA_C3-top	ACACCGCAGATGTAGTGTTCACAG
sgRNA_H1-top	ACACCCTGCCAACAGGGCAGTAAG
sgRNA_E1-top	ACACCGAGTCGGAGCAGAAGAAGAAG
sgRNA_V1-top	ACACCGGGTGGGGGAGTTGCTCCG
sgRNA_A1-bottom	ACACCGATGGAGCCAGAGAGGATCCG
sgRNA_C1-bottom	AAAACCAAGTCTAGCAAGCAGGCCAG
sgRNA_C3-bottom	AAAACTGTGAAACACTACATCTGCG
sgRNA_H1-bottom	AAAACTTCTTCTGCTCGGACTCG
sgRNA_E1-bottom	AAAACTTACTGCCCTGTGGGGCAAGG
sgRNA_V1-bottom	AAAACGGAGCAAACCCCCCACCG
PCR_Pla-fwd	AGG AAA GAA CAT GTG AGC AAA AG
PCR_Pla-rev	CAGCGAGTCAGTGAGCGA
PCR_sgRNA-fwd1	CTGTACAAAAAAGCAGGCTTTA
PCR_sgRNA-rev1	AACGTAGGTCTTACCGCTGTACAAAAAAGCAGGCTTTA AAAAAAAGCACCAGACTCGGTGCCACTTTCAAGTTGATAACGGACTAGCCTTAT
PCR_sgRNA-rev2	TTAACCTGCTATTCTAGCTAAAC TTGCTATTCTAGCTCTAAACTCTCTCCTGCCAGAACCGGTGTTCGTCCTT
PCR_sgRNA_A2	CCA TTGCTATTCTAGCTCTAAACCGCCGAGGTGAAGTTCGAGGGTGTTCGTCCTT
PCR_sgRNA_G1	TCCA TTGCTATTCTAGCTCTAAACAGGTGAAGTTCGAGGGCGACGGTGTTCGTCCTT
PCR_sgRNA_G2	TCCA TTGCTATTCTAGCTCTAAACTGGTGAACCGCATCGAGCTGGGTGTTCGTCCTT
PCR_sgRNA_G3	TCCA TTGCTATTCTAGCTCTAAACTCAGCTCGATGCCGTTACCGGTGTTCGTCCTT
PCR_sgRNA_G4	TCCA TTGCTATTCTAGCTCTAAACTCAGCTCGATGCCCTCAGCGGTGTTCGTCCTT
PCR_sgRNA_G5	TCCA TTGCTATTCTAGCTCTAAACTCAGCTCGATGCCGTTACCGGTGTTCGTCCTT
PCR_sgRNA_G6	CCA TTGCTATTCTAGCTCTAAACGGATGTTGCCGTCCTGGGTGTTCGTCCTT
PCR_sgRNA_G7	CCA TTGCTATTCTAGCTCTAAACGCTTGAGGGAGATGAGGACTGGTGTTCGTCCTT
PCR_sgRNA_C2	TCCA TTGCTATTCTAGCTCTAAACATGACTGTGAAGAGCTTCACGGTGTTCGTCCTT
PCR_sgRNA_C4	TCCA TTGCTATTCTAGCTCTAAACGAGGACAAAGTACAAACGGCGGTGTTCGTCCTT
PCR_sgRNA_E2	TTCCA TTGCTATTCTAGCTCTAAACGAACGGAGGACAAAGTACAGGTGTTCGTCCTT
PCR_sgRNA_E3	TTCCA TTGCTATTCTAGCTCTAAACACCACCAACTTCATCCACGGGTGTTCGTCCTT
PCR_sgRNA_H2	TCCA TTGCTATTCTAGCTCTAAACGGGCCTCACCAACTTCAGGTGTTCGTCCTT
PCR_sgRNA_H3	TCCA TTGCTATTCTAGCTCTAAACGCCAGGGCCTCACCAAGGTGTTCGTCCTT
PCR_sgRNA_H4	TCCA TTGCTATTCTAGCTCTAAACACCTGCCAGGGCCTCACCAAGGTGTTCGTCCTT
PCR_sgRNA_H5	TCCA TTGCTATTCTAGCTCTAAACTGATAACCAACCTGCCAGGGGTGTTCGTCCTT
PCR_sgRNA_H6	TCCA

PCR_sgRNA_H7	TTGCTATTCTAGCTCTAAACTAACCTGTCTGTAACCTGGTTCGTCCTT CCA
PCR_sgRNA_V2	TTGCTATTCTAGCTCTAAAACGCTCTGGCTAAAGAGGGAATGGTTCGTCCTT TCCA
PCR_sgRNA_V3	TTGCTATTCTAGCTCTAAAACCGGCTCTGGCTAAAGAGGGAGGTTCGTCCTT TCCA
PCR_sgRNA_V4	TTGCTATTCTAGCTCTAAAACCTCTGCACACCCGGCTGGGGTTCGTCCTT TCCA
PCR_sgRNA_SA1	TTGCTATTCTAGCTCTAAAACCTCTGAGCCTCAGTTCGGTTCGTCCTT CCA
PCR_sgRNA_SA2	TTGCTATTCTAGCTCTAAAACGCATGTCCATTCCACCTTAGGTTCGTCCTT CCA
PCR_sgRNA_SB1	TTGCTATTCTAGCTCTAAAACCTGTGCCTGGCTCTAATTGGTTCGTCCTT CCA
PCR_sgRNA_SB2	TTGCTATTCTAGCTCTAAAACCCACTGTACAATGTTGCAGGTTCGTCCTT TCCA
PCR_sgRNA_SC1	TTGCTATTCTAGCTCTAAAACCGTGCCTGGCTCAAATGCGGTTCGTCCTT TCCA
PCR_sgRNA_SC2	TTGCTATTCTAGCTCTAAAACAGTTGTGAGGAAAGCAGTGGGTTCGTCCTT TCCA
Survey_GFP-fwd	TACGGCAAGCTGACCTGAA
Survey_GFP-rev	GTCCATGCCGAGAGTGATCC
Surveye_CLTA-fwd	GCCAGGGCTGTTATCTTGG
Surveye_CLTA-rev	ATGCACAGAACAGCAGGTTGA
Survey_EMX-fwd	CTGTGTCCTCTCCTGCCCT
Survey_EMX-rev	CTCTCCGAGGAGAACGCCAA
Survey_HBB-fwd	GGTAGACCACCAGCAGCCTA
Survey_HBB-rev	CAGTGCCAGAACAGCCAAGG
Survey_VEGF-fwd	CCACACAGCTTCCCCTCTC
Survey_VEGF-rev	GAGAGCCGTTCCCTCTTGC
Survey_AAVS1-fwd	ACTGCTTCTCCTCTGGGAAGT
Survey_AAVS1-rev	TCATGGCATCTTCCAGGGGT
HTS_EXM_ON-fwd	CACTTTCCCTACACGACGCTCTCCGATCT CCTCCCCATTGGCCTGCTTC
HTS_EXM_Off1-fwd	CACTTTCCCTACACGACGCTCTCCGATCT TCGTCCTGCTCTCACTTAGAC
HTS_EXM_Off2-fwd	CACTTTCCCTACACGACGCTCTCCGATCT TTTGTGGCTGGCCCCAGT
HTS_EXM_Off3-fwd	CACTTTCCCTACACGACGCTCTCCGATCT TGCACTGCTCATGACTGGCCT
HTS_EXM_Off4-fwd	CACTTTCCCTACACGACGCTCTCCGATCT TTCTGAGGGCTGCTACCTGT
HTS_VEFG_ON-fwd	CACTTTCCCTACACGACGCTCTCCGATCT ACATGAAGCAACTCCAGTCCC
HTS_EXM_Off1-fwd	CACTTTCCCTACACGACGCTCTCCGATCT AGCAGACCCACTGAGTCAACTG
HTS_EXM_Off2-fwd	CACTTTCCCTACACGACGCTCTCCGATCT CCCGCCACAGTCGTGTCAT
HTS_EXM_Off3-fwd	CACTTTCCCTACACGACGCTCTCCGATCT CGCCCCGGTACAAGGTGA
HTS_EXM_Off4-fwd	CACTTTCCCTACACGACGCTCTCCGATCT GTACCGTACATTGAGGATGTT
HTS_CLTA2_ON-fwd	CACTTTCCCTACACGACGCTCTCCGATCT CCTCATCTCCCTCAAGCAGGC
HTS_CLTA2_Off1-fwd	CACTTTCCCTACACGACGCTCTCCGATCT ATTCTGCTCTGAGGTTATTGTT
HTS_CLTA2_Off2-fwd	CACTTTCCCTACACGACGCTCTCCGATCT
HTS_CLTA2_Off3-fwd	CACCTCTGCCTCAAGAGCAGAAAA
HTS_EXM_ON-rev	CACTTTCCCTACACGACGCTCTCCGATCT TGTGTGTGTGTGTAGGACT GGAGTTCAGACGTGTGCTCTCCGATCT TCATCTGCCCCCTCCCTCC

HTS_EXM_Off-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CGAGAAGGAGGTGCAGGAG
HTS_EXM_Off-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CGGGAGCTGTCAGAGGCTG
HTS_EXM_Off-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CTCACCTGGCGAGAAAGGT
HTS_EXM_Off-rev	GGAGTTCAGACGTGTGCTCTCCGATCT AAAACTCAAAGAAATGCCAATCA
HTS_VEFG_ON-rev	GGAGTTCAGACGTGTGCTCTCCGATCT AGACGCTGCTCGCTCCATTCA
HTS_EXM_Off1-rev	GGAGTTCAGACGTGTGCTCTCCGATCT ACAGGCATGAATCACTGCACCT
HTS_EXM_Off2-rev	GGAGTTCAGACGTGTGCTCTCCGATCT GCGGCAACTTCAGACAACCGA
HTS_EXM_Off3-rev	GGAGTTCAGACGTGTGCTCTCCGATCT GACCCAGGGGACCAGTT
HTS_EXM_Off4-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CTGCCTTCATTGCTTAAAAGTGGAT
HTS_CLTA2_ON-rev	GGAGTTCAGACGTGTGCTCTCCGATCT ACAGTTGAAGGAAGGAAACATGC
HTS_CLTA2_Off1-rev	GGAGTTCAGACGTGTGCTCTCCGATCT GCTGCATTGCCATTCCA
HTS_CLTA2_Off2-rev	GGAGTTCAGACGTGTGCTCTCCGATCT GTTGGGGAGGAGGAGCTTAT
HTS_CLTA2_Off3-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CTAAGAGCTATAAGGGCAAATGACT
HTS_SiteA_On-fwd	CACTTTCCCTACACGACGCTCTCCGATCT AATCCCAGCTTGCCACCAC
HTS_SiteA_Off1-fwd	CACTTTCCCTACACGACGCTCTCCGATCT GTGCATTGAGGAGATGAACCAGC
HTS_SiteA_Off2-fwd	CACTTTCCCTACACGACGCTCTCCGATCT AAGACAGATGCCAGTCCCT
HTS_SiteA_Off3-fwd	CACTTTCCCTACACGACGCTCTCCGATCT CGGGTCGAATCCTGATGCC
HTS_SiteB_On-fwd	CACTTTCCCTACACGACGCTCTCCGATCT CTCGGCTTCCAAACTGCTG
HTS_SiteB_Off1-fwd	CACTTTCCCTACACGACGCTCTCCGATCT ATCCACCCACCTCAGCCTCT
HTS_SiteB_Off2-fwd	CACTTTCCCTACACGACGCTCTCCGATCT TCGACCTCCAAAGTGCTGG
HTS_SiteB_Off3-fwd	CACTTTCCCTACACGACGCTCTCCGATCT GCCTCCAAAGTGCTGGGAT
HTS_SiteB_Off4-fwd	CACTTTCCCTACACGACGCTCTCCGATCT ACCCGCTTCAGCCTCC
HTS_SiteC_On-fwd	CACTTTCCCTACACGACGCTCTCCGATCT AATCCTCCGCCTCAGCTTC
HTS_SiteC_Off1-fwd	CACTTTCCCTACACGACGCTCTCCGATCT GATCCGCCTGTCGACCTC

HTS_SiteC_Off2-fwd	CACTCTTCCCTACACGACGCTCTCCGATCT GAGCTCAGGCAATCTGCCA
HTS_SiteC_Off3-fwd	CACTCTTCCCTACACGACGCTCTCCGATCT GGCCTCCAAAGTGCTGGTA
HTS_SiteC_Off4-fwd	CACTCTTCCCTACACGACGCTCTCCGATCT AGTGATCCACCTGCCTCAGC
HTS_SiteA_On-rev	GGAGTTCAGACGTGTGCTCTCCGATCT GAACATGCCCTGGTAGGGGG
HTS_SiteA_Off1-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CCATCAGCACCAAAAGGCA
HTS_SiteA_Off2-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CAGCTGCATATCCAGTGGC
HTS_SiteA_Off3-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CCTGTTCCATGCCACAGACT
HTS_SiteB_On-rev	GGAGTTCAGACGTGTGCTCTCCGATCT AGCCAGACAGACTATGCCCT
HTS_SiteB_Off1-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CGGGGGACAGAAGGGAAGG
HTS_SiteB_Off2-rev	GGAGTTCAGACGTGTGCTCTCCGATCT ACTCAACATACAAAAATCAATGGTG
HTS_SiteB_Off3-rev	GGAGTTCAGACGTGTGCTCTCCGATCT TTCTGGCATCTGGCCTCTGG
HTS_SiteB_Off4-rev	GGAGTTCAGACGTGTGCTCTCCGATCT ATCCACCCGCTTGGC
HTS_SiteC_On-rev	GGAGTTCAGACGTGTGCTCTCCGATCT AGTTCAATTGCATTCACAGTTCCA
HTS_SiteC_Off1-rev	GGAGTTCAGACGTGTGCTCTCCGATCT CACAGCCACCACCAACTG
HTS_SiteC_Off2-rev	GGAGTTCAGACGTGTGCTCTCCGATCT ACAAGTAGAGAACAGAGAAGTGAA
HTS_SiteC_Off3-rev	GGAGTTCAGACGTGTGCTCTCCGATCT GTCTAATGAAGTTGCTAATAGAGGAA
HTS_SiteC_Off4-rev	GGAGTTCAGACGTGTGCTCTCCGATCT TCCTCCCACTTCAGCCTTCA

FokI and Cas9 Protein Domain Sequences

Cas9:

MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAETR
 LKRTARRRYTRRKNRICYLQEIFSNEAKVDDSSFFHRLEESFLVEEDKKHERHPIFGNIVDEVA
 YHEKYPTIYHLRKKLVDSTDKAIDLRLIYLALAHMIKFRGHFLIEGDLNPNSDVKLFQLVQ
 TYNQLFEENPINASGVDAKAILSARLSKSRRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSN
 FDLAEDAKLQLSKDTYDDDDLNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLS
 ASMIKRYDEHHQDLTLLKALVRQQLPEKYKEIFFDQSCKNGYAGYIDGGASQEEFYKFIKPILEK
 MDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYPFLKDNRREKIEKILTFR
 IPYYVGPLARGNSRAWMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKLPNEKVLPK
 HSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKI
 ECFDSVEISGVVEDRFNASLGYHDLLKIIKDKDFLDNEENEDILEDIVLTTLFEDREMIEERLK
 TYAHLFDDKVMKQLKRRRTGWGRRLSRKLINGIRDQSGKTILDFLKSDGFANRNFMQLIH

DSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAICKGILQTVKVVDELVKVMGRHKPENIVIE
MARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYLQNGRDMY
VDQELDINRLSDYDWDHVIPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKNYWRQ
LLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQILDLSRMNTKYDENDK
LIREVKVITLKSCLVSDFRKDFQFYKREINNYHHADAYLNAVVTALIKKYPKLESEFVYG
DYKVYDVRKMIAKSEQEIGKATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWD
KGRDFATVRKVLMPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWPCKYGGFDSP
TVAYSVLVVAKVEKGSKKLKSVKELLGITMERSSFEKNPIDFLEAKGYKEVKKDLIILPKY
SLFELENGRKMLASAGELQKGNELALPSKYVNFLYASHYEKLKGSPEDNEQKQLFVEQHK
HYLDEIIEQISEFSKRVILADANLDKVLSAYNHRDKPIREQAENIIHLFTLTNLGAPAAFKYFD
TTIDRKRYTSTKEVLDATLIHQSITGLYETRIDLSQLGGD

Cas9 nickase (**D10A**):

MDKKYSIGL~~A~~IGTNSVGWAVITDEYKVPSSKKFVLGNTDRHSIKKNLIGALLFDSETAEATR
LKRTARRRYTRRKNRICYLQEIFSNEAKVDDSFHRLEESFLVEEDKKHERHPIFGNIVDEVA
YHEKYPTIYHLRKKLVDSTDKAIDLRLIYLALAHMIKFRGHFLIEGDLNPDSVDKLFQLVQ
TYNQLFEENPINASGVDAKAILSARLSKSRRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSN
FDLAEDAKLQLSKDTYDDLDNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLS
ASMIKRYDEHHQDLTLLKALVRQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEK
MDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYPFLKDNREKIEKILTFR
IPYYVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSIFIERMTNFDKLPNEKVLPK
HSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKI
ECFDSVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTTLFEDREMIEERLK
TYAHLFDDKVMKQLKRRRTGWRGLSRKLINGIRDQSGKTILDFLSDGFANRNFMQLIHD
DSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAICKGILQTVKVVDELVKVMGRHKPENIVIE
MARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYLQNGRDMY
VDQELDINRLSDYDWDHVIPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKNYWRQ
LLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQILDLSRMNTKYDENDK
LIREVKVITLKSCLVSDFRKDFQFYKREINNYHHADAYLNAVVTALIKKYPKLESEFVYG
DYKVYDVRKMIAKSEQEIGKATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWD
KGRDFATVRKVLMPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWPCKYGGFDSP
TVAYSVLVVAKVEKGSKKLKSVKELLGITMERSSFEKNPIDFLEAKGYKEVKKDLIILPKY
SLFELENGRKMLASAGELQKGNELALPSKYVNFLYASHYEKLKGSPEDNEQKQLFVEQHK
HYLDEIIEQISEFSKRVILADANLDKVLSAYNHRDKPIREQAENIIHLFTLTNLGAPAAFKYFD
TTIDRKRYTSTKEVLDATLIHQSITGLYETRIDLSQLGGD

dCas9 (**D10A and H840A**):

MDKKYSIGL~~A~~IGTNSVGWAVITDEYKVPSSKKFVLGNTDRHSIKKNLIGALLFDSETAEATR
LKRTARRRYTRRKNRICYLQEIFSNEAKVDDSFHRLEESFLVEEDKKHERHPIFGNIVDEVA
YHEKYPTIYHLRKKLVDSTDKAIDLRLIYLALAHMIKFRGHFLIEGDLNPDSVDKLFQLVQ
TYNQLFEENPINASGVDAKAILSARLSKSRRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSN
FDLAEDAKLQLSKDTYDDLDNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLS
ASMIKRYDEHHQDLTLLKALVRQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEK
MDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYPFLKDNREKIEKILTFR
IPYYVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSIFIERMTNFDKLPNEKVLPK
HSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKI
ECFDSVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTTLFEDREMIEERLK
TYAHLFDDKVMKQLKRRRTGWRGLSRKLINGIRDQSGKTILDFLSDGFANRNFMQLIHD

DSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKGILQTVKVVDELVKVMGRHKPENIVIE
MARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYLQNGRDMY
VDQELDINRLSDYDVA**A**IVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKNYWRQ
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DNA Coding Sequences of wild-type Cas9 Nuclease, Cas9 Nickase and *FCas9* fusions

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dCas9-NLS-GGS3linker-*FokI*:

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NLS- dCas9-GGS3linker -*FokI*:

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